

DIVISIONAL APPLICATION
FOR
UNITED STATES LETTERS PATENT

TITLE: DWF4 POLYNUCLEOTIDES, POLYPEPTIDES AND USES
THEREOF

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DWF4 POLYNUCLEOTIDES, POLYPEPTIDES AND USES THEREOF

5 CROSS-REFERENCE TO RELATED APPLICATIONS

This application is related to provisional patent applications serial nos. 60/119,657, filed February 11, 1999 and 60/119,658, filed February 11, 1999, from which priority is claimed under 35 USC §119(e)(1) and which applications are incorporated herein by reference in their entireties.

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TECHNICAL FIELD

The present invention relates to novel polynucleotides isolated from dwarf plants. The *dwf4* polynucleotides encode all, or a portion of, a DWF4 polypeptide, a cytochrome P450 enzyme that mediates multiple steps in synthesis of
15 brassinosteroids. The present invention also relates to isolated polynucleotides that encode regulatory regions of *dwf4*. Uses of the *dwf4* polypeptides and polynucleotides are also disclosed.

BACKGROUND

20 Plant growth is accomplished by orderly cell division and tightly regulated cell expansion. In plants, the contribution of cell expansion to growth is of much greater significance than in most other organisms; all plant organs owe their final size to a period of significant cell elongation, which usually follows active cell division. Further, the sessile nature of plants requires that they make fine but responsive
25 adjustments in growth to survive harsh environmental conditions and to optimize their use of limited resources (Trewavas (1986) "Resource allocation under poor growth conditions: A major role for growth substances in developmental plasticity" In Plasticity in Plants, D.H. Jennings and A.J. Trewavas, eds (Cambridge, UK: Company of Biologists Ltd.), pp. 31-76).

30 In Arabidopsis, cell elongation is largely responsible for hypocotyl growth in germinating seedlings and extension of inflorescences (bolting) at the end of vegetative growth. Coordinate control of plant growth is regulated by both external

stimuli and internal mechanisms. Of the external signals, the most obvious is light (Deng, X.-W. (1994) *Cell* 76:423-426). Light inhibits hypocotyl elongation and promotes cotyledon expansion and leaf development in seedlings, and photoperiod is crucial for flower initiation in a large number of species.

5 The internal components of plant signaling are generally mediated by chemical growth regulators (phytohormones; reviewed in Klee, H., and Estelle, M. (1991) *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 42:529-551). Thus, plant growth in response to environmental factors is modulated by plant hormones acting alone or in concert (Evans "Functions of hormones at the cellular level of organization" In
10 Hormonal Regulation of Plant Physiology, T.K. Scott, ed (Berlin: Springer-Verlag), pp. 23-79), and growth depends on regulated cellular events, such as division, elongation, and differentiation.

 Gibberellic acid (GA) and cytokinins promote flowering; in addition, GA stimulates stem elongation, whereas cytokinins have the opposite effect, reducing
15 apical dominance by stimulating increased axillary shoot formation. Conversely, auxins promote apical dominance and stimulate elongation by a process postulated to require acidification of the cell wall by a K⁺-dependent H⁺-pumping ATPase (Rayle, D.L., and Cleland, R.E. (1977) *Curr. Top. Dev. Biol.* 11:187-214).

 In addition to the classic hormones, such as auxin and gibberellic acid (GA),
20 brassinosteroids (BRs) have been discovered to be important in growth promotion (reviewed in Clouse (1996) *Plant J.* 10:1-8). The most recently discovered class of plant growth substances, the BRs, has been to date the least studied; however, rapid progress toward understanding BR biosynthesis and regulation is now being made (Yokota, T. (1997) *Trends Plant Sci.* 2:137-143). The term BRs collectively refers to
25 the growth-promoting steroids found in plants (Grove et al. (1979) *Nature* 281:216-217). They are structurally very similar to the molting hormones of insects, ecdysteroids (Richter and Koolman (1991) "Antiecdysteroid effects of brassinosteroids in insects" in *Brassinosteroids: Chemistry, Bioactivity, and Applications*, H.G. Cutler, T. Yokota, and G. Adam, eds (Washington, DC: American
30 Chemical Society), pp. 265-279), but active BRs have unique structural features. As shown in Figure 1, a 6-oxolactone or 7-oxalactone in the B ring, 5 α hydrogen, and multiple hydroxylations at four different positions with specific stereochemistry have

been proposed as an essential configuration for BRs (reviewed in Marquardt and Adam (1991) "Recent advances in brassinosteroid research" in Chemistry of Plant Protection, W. Ebing, ed (Berlin: Springer-Verlag), pp. 103-139). Among >40 naturally occurring BRs, brassinolide (BL; 2 α , 3 α , 22(R), 23(R)-tetrahydroxy-24(S)-methyl-B-homo-7-oxa-5 α -cholestan-6-one) has been shown to be the most biologically active (reviewed in Mandava (1988) *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 39:23-52). As a major biological effect, BRs stimulate longitudinal growth of young tissues via cell elongation and cell division (reviewed in Clouse (1996), *supra*; Fujioka and Sakurai (1997a) *Nat. Prod. Rep.* 14:1-10).

Elucidating the BR biosynthetic pathways has been a major area of recent interest. Biochemical analyses have been used to elucidate the BR biosynthetic pathway (Fujioka et al. (1996) *Plant Cell Physiol.* 37:1201-1203; Choi et al. (1997), *Phytochemistry* 44:609-613), and mutational analyses are being used to confirm this pathway. Similar to the biosynthetic pathways of the human steroid hormones and insect ecdysteroids (Rees (1985) "Biosynthesis of ecdysone" in Comprehensive Insect Physiology, Biochemistry and Pharmacology, G.A. Kerkut and L.I. Gilbert, eds (Oxford, UK: Pergamon Press), pp. 249-293; Granner, D.K. (1996) "Hormones of the gonads" in Harper's Biochemistry, R.K. Murray, D.K. Granner, P.A. Mayes, and V.W. Rodwell, eds (Stamford, CT: Appleton and Lange Press), pp. 566-580), BRs are synthesized via multiple parallel pathways (Fujioka et al. (1996) *Plant Cell Physiol.* 37:1201-1203; Choi et al. (1997), *supra*). Starting from the initial precursor, campesterol (CR), the BR intermediates undergo a series of hydroxylations, reductions, an epimerization, and a Baeyer-Villiger-type oxidation leading to the most oxidized form, BL (Fujioka and Sakurai (1997b) *Physiol. Plant.* 100:710-715; Figure 1). Castasterone (CS) oxidation, the last step in BR biosynthesis, is not found in some species, such as mung bean. In that case, CS plays a role as the major BR rather than BL (Yokota et al. (1991) "Metabolism and biosynthesis of brassinosteroids" in Brassinosteroids: Chemistry, Bioactivity, and Application, H.G. Cutler, T. Yokota, and G. Adam, eds (Washington, DC: American Chemical Society), pp. 86-96). Traditionally, BR biosynthetic pathways have been elucidated by feeding deuterio-labeled intermediates to BR-producing cell lines of Madagascar periwinkle (Sakurai and Fujioka (1996) "Catharanthus roseus (Vinca rosea): In vitro production

of brassinosteroids" in *Biotechnology in Agriculture and Forestry*, Y.P.S. Bajaj, ed (Berlin: Springer-Verlag), pp. 87-96.). The present model, including parallel branched pathways and early and late C-6 oxidation pathways, was established using these feeding studies (Fujioka and Sakurai (1997a), *supra*, Fujioka and Sakurai
5 (1997b), *supra*; Sakurai and Fujioka (1997) *Biosci. Biotechnol. Biochem.* 61:757-762).

Although the brassinosteroid system is a less well understood class of plant growth substances (BRs; Mitchell, et al. (1970) *Nature* 225:1065-1066; Grove et al. (1979) *Nature* 281:216-217; Mandava, N.B. (1988) *Annu. Rev. Plant Physiol. Plant
10 Mol. Biol.* 39:23-52), several such compounds have been identified and are known to effect elongation of cells in various plant tissues, their biosynthesis, regulation, and mechanism of action have only recently begun to be elucidated (reviewed in Clouse, S.D. (1996) *Plant J.* 10:1-8; Fujioka, S., and Sakurai, A. (1997) *Physiol. Plant.* 100:710-715).

15 Several types of dwarf or dwarflike mutants have been described in *Arabidopsis*. A number of mutations have been identified that affect either light-dependent (*cop*, *det*, and *fusca* [fus; another group of mutants with some members perturbed in light-regulated growth]) or hormone signaling (*axr2*) pathways and whose pleiotropic phenotypes include defects in cell elongation. The majority of
20 these mutants also have other alterations in their phenotypes. At least five GA mutants have been described as being reduced in stature (Koornneef and Van der Veen (1980) *Theor. Appl. Genet.* 58:257-263). GA biosynthetic mutants may also have no or defective flower development and are marked by an absence of viable pollen. Reduced levels of endogenous gibberellins are also a characteristic (Barendse
25 et al.(1986) *Physiol. Plant.* 67:315-319; Talon et al. (1990) *Proc. Natl. Acad. Sci. USA* 87:7983-7987), and their phenotype can be nearly restored to that of the wild type by the addition of exogenous GA. (Koornneef and Van der Veen (1980) *Theor. Appl. Genet.* 58:257-263).

Another hormone mutation, *auxin resistant2* (*axr2*), results in plants with a
30 dwarf phenotype both in the light and in darkness as well as increased resistance to high levels of auxin, ethylene, and abscisic acid (Timppe et al. (1992) *Planta* 188:271-278). An interesting relationship exists between light regulation and

cytokinin levels. Arabidopsis seedlings grown in the dark in the presence of cytokinins have open cotyledons, initiate chloroplast differentiation and leaf development, and activate transcription from the chlorophyll *a/b* binding protein gene (*CAB*) promoter. Importantly, they also display a cytokinin dose-dependent dwarf
5 phenotype.

Dwarf Arabidopsis mutants that are rescued by addition of BRs have also been described (Kauschmann et al. (1996) *Plant J.* 9:701-713; Li et al. (1996) *Science* 272:398-401; Szekeres et al. (1996) *Cell* 85:171-182; Azpiroz et al. (1998) *Plant Cell* 10:219-230), including the following three mutants: dwarf1 (*dwf1*; Kauschmann et al.
10 (1996) *Plant J.* 9:701-713), constitutive photomorphogenesis and dwarfism (*cpd*; Szekeres et al. (1996) *Cell* 85:171-182), and *det2* (Li et al. (1996) *Science* 272:398-401). These mutants have been shown to be defective in steroid biosynthesis. *DWF1* (Feldmann et al. (1989) *Science* 243:1351-1354) was cloned first (GenBank accession number U12400). Takahashi et al. (1995) *Genes Dev.*
15 9:97-107 hypothesized that *DWF1*, which they isolated with an allele of *dwf1*, referred to as diminuto1 (*dim1*), contains a potential nuclear targeting signal, which may confer a regulatory function to the protein. However, Mushegian and Koonin (1995) *Protein Sci.* 4:1243-1244 indicated that *DWF1* displays limited homology with flavin adenine dinucleotide (FAD)independent oxidoreductase, suggesting an
20 enzymatic function in BR biosynthesis. According to Kauschmann et al. (1996), *supra* (*dwf1-6* described as *cabbage1* [*cbb1*]), *dwf1* mutants were rescued by exogenous application of BRs.

DET2 was shown to encode a putative steroid 5 α -reductase, mediating an early step in BR biosynthesis (Li et al. (1996), *supra*, Li et al. (1997) *Proc. Natl.*
25 *Acad. Sci. USA* 94:3554-3559; Fujioka et al. (1997) *Plant Cell* 9:1951-1962; Figure 1). Moreover, *det1* and *det2* have a decreased requirement for cytokinins in tissue culture and appear to be saturated for a cytokinin-dependent delay in senescence (Chory et al. (1994) *Plant Physiol.* 104:339-347). *CPD* has been proposed to be a novel cytochrome P450 (CYP90A1; Szekeres et al. (1996), *supra*), encoding a
30 putative 23 α -hydroxylase that acts in BR biosynthesis. The range of phenotypes in the *deetiolated* (*det*) and *constitutive photomorphogenic* (*cop*) light-regulatory mutants is broad. Mutations in *DET1*, *COP1*, *COP8*, *COP9*, *COP10*, and *COP11*

result in constitutive derepression of substantial portions of the photomorphogenic program (Chory, et al. (1989b) *Cell* 58:991-999; Deng, X.-W., and Quail, P.H. (1992) *Plant J.* 2:83-95; Wei, N., and Deng, X.-W. (1992) *Plant Cell* 4:1507-1518; Wei et al. (1994) *Plant Cell* 6:629-643), whereas mutations in *COP4* seem to affect only

- 5 morphology and gene expression (Hou et al. (1993) *Plant Cell* 5:329-339). The only invariant phenotype in this class of light-regulatory mutants is a substantial reduction in height in both light and darkness.

- There are additional dwarfs that are insensitive to one of these hormones, such as *bri* (*brassinosteroid insensitive*; Clouse et al. (1996) *Plant Physiol.* 111:671-678; Li and Chory (1997) *Cell* 90:929-938), *gai* (*gibberellic acid insensitive*; Koornneef et al. (1985) *Physiol. Plant.* 65:33-39), and *axr2* (*auxin resistant2*; Timpte et al. (1994) *Genetics* 138:1239-1249). Clouse et al. (1996), *supra* isolated *bri* by screening ethyl methanesulfonate-mutagenized populations for mutants whose root growth is not retarded at inhibitory concentrations of BR. Thus, the BRI protein is proposed to be
- 15 involved in BR signal perception or transduction (Clouse (1996), *supra*).

- Kauschmann et al. (1996), *supra* described a phenotypically similar mutant *cbb2* that maps to the same location. In addition, the *dwf2* alleles possess a phenotype similar to *bri* and map to the same region (Feldmann and Azpiroz (1994) "*dwarf (dwf)* and *twisted dwarf (twd)*" in *Arabidopsis: An Atlas of Morphology and Development*, J. Bowman, ed (New York: Springer-Verlag), pp. 82-85). It seems likely that all of the BR-insensitive dwarf mutants described to date are allelic. Recently, *BRI* has been cloned and shown to encode a leucine-rich-repeat receptor kinase, suggesting a role in the BR signal transduction pathway (Li and Chory (1997), *supra*).
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- Mutants defective in BR biosynthesis have also been isolated in other plant species. Bishop et al. (1996) *Plant Cell* 8:959-969 isolated a tomato *dwarf* mutant by transposon tagging. The tomato *Dwarf* gene encodes a pioneering member of the CYP85 family, and it appears to be involved in BR biosynthesis. In addition, Nomura et al. (1997) *Plant Physiol.* 113:31-37 reported that the *lka* and *lkb* mutants in garden pea are deficient in BR biosynthesis (*lkb*) or perception (*lka*).
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- Currently, little is known about the downstream events that occur in response to these signals and thereby directly control cell size. This is because the biochemical and cell biological processes involved have thus far been difficult to address. In
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addition, there is little information about the integration of regulatory signals converging at the cell from different signaling pathways and the ways they are coordinately controlled. In particular, the interaction of light and hormones in the control of cell elongation is not clear. Thus, there remains a need for the
5 identification and characterization of additional mutants and polypeptides encoded thereby involved in these pathways of plant growth.

SUMMARY OF THE INVENTION

In one aspect the invention includes an isolated *dwf4* polynucleotide
10 comprising an open reading frame that encodes a polypeptide comprising (i) a sequence having greater than 43% identity to the amino acid sequence of SEQUENCE ID NO:2; (ii) a sequence comprising at least about 10 contiguous amino acids that have greater than 43% identity to 10 contiguous amino acids of SEQUENCE ID NO:2, or a complement or reverse complement of said
15 polynucleotide. In certain embodiments, the polynucleotide will have at least 70% identity to the DWF4 polypeptide-coding region of SEQ ID NO:1 or to complements and reverse complements of this region. In further embodiments, the isolated *dwf4* polynucleotide comprises the nucleotide sequence of SEQ ID NO:1, complements and reverse complements thereof. The polynucleotide may also comprise at least 30
20 consecutive nucleotides of SEQ ID NO:1.

In another aspect, the invention includes an isolated *dwf4* polynucleotide comprising (i) a sequence having at least 50% identity to SEQ ID NO:1, complements and reverse complements thereof or (ii) a sequence comprising at least about 15 contiguous nucleotides that has at least 50% identity to SEQ ID NO:1, complements
25 and reverse complements thereof. In certain embodiments, the isolated *dwf4* polynucleotide has at least 50% identity to the DWF4 polypeptide-coding region of SEQ ID NO:1, complements and reverse complements thereof. In further embodiments, the isolated *dwf4* polynucleotides described herein comprise the nucleotide sequence of SEQ ID NO:1, complements and reverse complements thereof
30 or nucleotide sequences comprising at least 30 consecutive nucleotides of SEQ ID NO:1. Any of the *dwf4* polynucleotides described herein may be genomic DNA and may include introns. Further, in other embodiments, the *dwf4* polynucleotide includes

a *dwf4* control element comprising a polynucleotide selected from the group consisting of (i) a sequence having at least 50% identity to nucleotides 1 to 3202 of SEQ ID NO:1; (ii) a fragment of (i) which includes a *dwf4* control element; and (iii) complements and reverse complements of (i) or (ii). In still further embodiments, the polynucleotide includes a *dwf4* control element comprising a polynucleotide selected from the group consisting of (i) a sequence having at least 50% identity to nucleotides 6111 to 6468 corresponding to the 3' UTR of SEQ ID NO:1; (ii) a fragment of (i) which includes a *dwf4* 3' UTR; and (iii) complements and reverse complements of (i) or (ii). In certain embodiments, the polynucleotide includes a *dwf4* polynucleotide selected from the group consisting of (i) a sequence having at least 50% identity to the sequences corresponding to the introns of SEQ ID NO:1; (ii) a fragment of (i) which includes a *dwf4* intro; and (iii) complements and reverse complements of (i) and (ii). Introns are found, for example, in the following regions: nucleotides 3424 to 3503 of SEQ ID NO:1; nucleotides 3829 to 3913 of SEQ ID NO:1; nucleotides 4067 to 4164 of SEQ ID NO:1; nucleotides 4480 to 4531 of SEQ ID NO:1; nucleotides 4725 to 4815 of SEQ ID NO:1; nucleotides 4895 to 5000 of SEQ ID NO:1; and nucleotides 5111 to 5864 of SEQ ID NO:1. 54. In still further embodiments, any of the polynucleotides described herein can operably linked to a nucleic acid molecule encoding a heterologous polypeptide (e.g., a cytochrome P450 polypeptide), for example, as a chimeric polynucleotide.

In another aspect, the invention includes recombinant vectors comprising (i) one or more of the polynucleotides described above; and (ii) control elements operably linked to the one or more polynucleotides, whereby a coding sequence within said polynucleotide can be transcribed and translated in a host cell. In certain embodiments, the recombinant vector comprises (a) any of the polynucleotides which include a *dwf4* control element described above (e.g., promoter or intron); and (b) a nucleic acid molecule comprising a coding sequence operably linked to the *dwf4* control element.

Host cells comprising and/or transformed with any of the recombinant vectors described herein are also provided. In certain embodiments, the host cells are cultured *ex vivo* while in other embodiments, the *dwf4* polynucleotide is provided the host cell

in vivo. In certain embodiments the DWF4 polypeptide is provided in amounts such that a plant is regenerated.

In another aspect, the present invention includes a method of modulating a DWF4 polypeptide comprising the following steps: (a) providing a host cell as
5 described herein; and (b) culturing said host cell under conditions whereby the *dwf4* polynucleotide included in the host cell is transcribed. In certain embodiments, the *dwf4* polynucleotide is overexpressed. Alternatively, in other embodiments, the polynucleotide included in the host cell inhibits expression of *dwf4*.

In yet another aspect, the present invention includes a transgenic plant
10 comprising any of the recombinant vectors described herein.

In yet another aspect, the invention includes a method of producing a recombinant polypeptide comprising the following steps: (a) providing a host cell as described herein; and (b) culturing said host cell under conditions whereby the recombinant polypeptide encoded by the coding sequence present in said recombinant
15 vector is expressed.

In a still further aspect, the invention includes a method of producing a transgenic plant comprising the steps of (a) introducing a polynucleotide described herein into a plant cell to produce a transformed plant cell; and (b) producing a transgenic plant from the transformed plant cell.

20 Methods for producing a transgenic plant having an altered phenotype relative to the wild-type plant comprising the following steps: introducing at least one polynucleotide described herein into a plant cell; and producing a transgenic plant from the plant cell, said transgenic plant having an altered phenotype relative to the wild-type plant are also included in the present invention. The altered phenotype
25 includes altered morphological appearance and altered biochemical activity, for example, altered (reduced or increased) cell length in any cell or tissue, altered (extended or decreased) periods of flowering, altered (increased or decreased) branching, altered (increased or decreased) seed production, altered (increased or decreased) leaf size, altered (elongated or shortened) hypocotyls, altered (increased or
30 decreased) plant height, altered heme-thiolate enzyme activity, altered monooxygenase activity, altered 22 α -hydroxylase activity, regulation of brassinosteroid synthesis, regulation of gibberellic acid, regulation of cytokinins,

regulation of auxins, altered resistance to plant pathogens, altered growth at low temperatures, altered growth in dark conditions and altered sterol composition. In certain embodiments, the at least one polynucleotide is operably linked to a promoter selected from the group consisting of a tissue-specific promoter, an inducible
5 promoter or a constitutive promoter. The polynucleotide can be overexpressed or it can inhibit expression of *dwf4*. In a still further embodiment, at least two polynucleotides are introduced into the plant cell. Each polynucleotide is operably linked to a different tissue-specific promoter such that one polynucleotide is overexpressed while the other inhibits expression of *dwf4*.

10 In yet another aspect, the invention includes a method for altering the biochemical activity of a cell comprising the following steps: introducing at least one polynucleotide described herein; and culturing the cell under conditions such that the biochemical activity of the cell is altered. Biochemical activity includes, for example, altered heme-thiolate enzyme activity, altered monooxygenase activity, altered 22 α -
15 hydroxylase activity, regulation of gibberellic acid, regulation of cytokinins, regulation of auxins, and altered sterol composition. In certain embodiments, the cell is cultured *ex vivo*. In other embodiments, the *dwf4* polynucleotide is provided to the cell *in vivo*. In still other embodiments, more than one *dwf4* polynucleotides are provided to the cell.

20 In yet another aspect, the invention includes a method for regulating the cell cycle of a plant cell comprising the following steps providing a *dwf4* polynucleotide to a plant cell; and expressing the *dwf4* polynucleotide to provide a DWF4 polypeptide, wherein the DWF4 polypeptide is provided in amounts such that cell cycling is regulated. In certain embodiments, the plant cell is provided *in vitro* and is
25 cultured under conditions suitable for providing the DWF4 polypeptide. In still other embodiments, the *dwf4* polynucleotide is provided *in vivo*.

In yet another aspect, the invention includes an isolated DWF4 polypeptide comprising (i) a sequence having greater than 43% identity to SEQ ID NO:2 or (ii) fragments of (i) that confer a DWF4 phenotype when expressed in a host organism.

30 In certain embodiments, the isolated DWF4 polypeptide comprises the amino acid sequence of SEQ ID NO:2. In certain embodiments, the invention includes a chimeric

polypeptide comprising a DWF4 polypeptide as described herein and a heterologous polypeptide, for example a cytochrome P450 polypeptide.

Any of the polynucleotides or polypeptides described herein can be used in diagnostic assays; to generate antibodies. Further, the antibodies and fragments thereof can also be used in diagnostic assays, to produce immunogenic compositions or the like.

These and other objects, aspects, embodiments and advantages of the present invention will readily occur to those of ordinary skill in the art in view of the disclosure herein.

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BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 depicts a proposed biosynthetic pathway for BL. CR goes through at least two different pathways, referred to as the early C-6 oxidation (right column) and late C-6 oxidation (left column) pathways. Steps mediated by DWF4, CPD
15 (Szerkeres et al. (1996), *supra*), DET2 (Fujioka and Skaurai (1997a), *infra*; Li et al. (1997), *supra*) and LKB (Yokota et al. (1997), *infra*) are indicated.

Figures 2A and B depict schematic representations of the *DWF4* gene and protein. Figure 2A depicts the *DWF4* coding sequence (1542 bp) and shows that the coding sequence contains eight exons and seven introns. The exons and introns range
20 in length from 93 to 604 and 84 to 754 bp, respectively. All of the introns are bordered by typical consensus splice junctions, 5'-GU and AG-3'. Closed rectangles indicate exons. The T-DNA position in *dwf4-1* is marked with an arrow. Figure 2B shows the relative positions of the major domains in DWF4 cytochrome P450. All of the major domains found in the cytochrome P450 superfamily are conserved in
25 DWF4. The estimated molecular mass and isoelectric point of the *DWF4* protein were 58 kD and 7.28, respectively. Hydropathy plotting and protein localization prediction by the PSORT software package (Nakai and Kaneshia (1992) *Genomics* 14:897-911) suggested that the protein may reside in a membrane of the endoplasmic reticulum as an integral protein. Mutations identified in the other *dwf4* alleles are
30 indicated.

Figure 3 depicts alignment of cytochrome P450 proteins that exhibited the most similarity to DWF4 in BLAST searches. GenBank accession numbers are

~~AF044216~~ (DWF4; CYP90B), ~~X87368~~ (CPD; CYP90A), ~~U54770~~ (tomato; CYP85),
~~D64003~~ (cyanobacteria; CYP120), ~~U32579~~ (maize; CYP88), ~~U68234~~ (zebrafish;
CYP26), and ~~M13785~~ (human; CYP3A3X). Dashes indicate gaps introduced to
maximize alignment. Domains indicated in Figure 2B are highlighted in a box. Amino
5 acid residues that are conserved >50% between the compared sequences are
highlighted by a reverse font, and identical residues between DWF4 and CPD are
boxed and italicized. Open triangles are placed under the 100% conserved residues.
Closed triangles locate functionally important amino acid residues, for example,
threonine (T) at 369, which is thought to bind molecular oxygen, and cysteine (C) at
10 516, which links to a heme prosthetic group by a thiolate bond. X's indicate mutated
residues in *dwf4* alleles. Multiple sequence alignment was performed using PILEUP
in the Genetics Computer Group package, and box shading was made possible by the
ALSCRIPT package (Barton (1993) *Protein Eng.* 6:37-40).

Figure 4 depicts the phylogenetic Relationship between DWF4 and Selected
15 Cytochrome P450s. DWF4 did not cluster with the group A plant cytochrome P450s
that are known to mediate plant-specific reactions (Durst and Nelson 1995). CYP90A,
CYP85, and DWF4, which are thought to be involved in BR metabolism, branched
from CYP88, which mediates GA biosynthesis. GenBank accession numbers for the
group A cytochrome P450s are M32885 (avocado; CYP71A1), P48421 (Arabidopsis;
20 CYP83), P48418 (petunia; CYP75A1), and X71658 (eggplant; CYP76A1). The
DISTANCE utility in the Genetics Computer Group software package was employed
to calculate the relationships.

Figure 5 depicts a comparison of wild-type and *dwf4* hypocotyl growth rates.
Circles indicate wild-type and square indicate *dwf4*. Each data point represents the
25 average of 10 seedlings.

Figure 6 depicts responses to cell elongation signals. BL measurements were
performed with *dwf4-3* and the corresponding wild-type control, Enkheim. Open bars
indicate the wild type. Filled bars indicate *dwf4*. Lines above the bars represent one
standard deviation. On the horizontal axis, "light" refers to light-grown controls;
30 "dark" refers to dark-grown controls; "hy2" refers to *DWF4* and *dwf4* plants in a *hy2*
background; "GA" refers to plants grown in 10^{-5} M GA; "2,4-D" refers to plants

grown in 10^{-8} M 2,4-D; "-BR" refers to liquid-grown controls; and "+BR" refers to liquid-grown controls with 10^{-8} M BL.

Figure 7 depicts pedicel elongation of *dwf4* mature plants in response to exogenous application of BR. Measurements were performed with the BR-fed plants. *dwf4-1* plants were more sensitive to intermediates belonging to the late C-6 oxidation pathway (10^{-6} M 6-deoxoCT and 10^{-6} M 6-deoxoTE) compared with compounds in the early C-6 pathway (10^{-5} M CT and 10^{-5} M TE). BL (10^{-7} M) induced almost the same amount of elongation with one-tenth the concentration of its precursors. Rescue by 22-OHCR (10^{-5} M), which is structurally similar to the presumed precursor CR, except for a 22 α -hydroxyl functional group, shows that the only defect in *dwf4* is the C-22 hydroxylation reaction. Complementing intermediates and BL induced dramatic elongation in the elongating zone of the inflorescence and pedicel, but fertility was not increased. Data represent the means \pm SE of 15 to 20 pedicels. "CTRL" refers to control; "WT" refers to wild type.

Figure 8 depicts the increase in inflorescence growth of three transformants which overexpress *dwf4* as compared to wild type (Ws-2). The length of inflorescences of DWF4 overexpression lines increased more than 20% compared to that of wild type. The length of the plant was measured at maturity. Each data point is a mean value of more than 9 plants, except AOD4-60 which represents 2 plants.

Figure 9 depicts the increase in seed production of three transformants which overexpress *dwf4* as compared to wild type (Ws-2). Seeds were harvested from individual plants of each genotype ($n > 5$). Seeds from each plant were weighed and a mean value calculated. The Figure shows percent increase over wild type.

Figures 10(A)-10(G) depict the nucleotide sequence of wild-type *dwf4* (SEQ ID NO:1, *see, also*, GenBank Accession Number AF044216). The *dwf4* polynucleotide includes a coding region between nucleotides 3203 and 6110, inclusive. The coding region includes the following eight exons: nucleotides 3203 to 3423, inclusive; nucleotides 3504 to 3828, inclusive; nucleotides 3914 to 4066, inclusive; nucleotides 4165 to 4479, inclusive; nucleotides 4632 to 4724, inclusive; nucleotides 4816 to 4894, inclusive; nucleotides 5001 to 5110, inclusive and nucleotides 5865 to 6110, inclusive. The exons are indicated by a bar beneath the nucleotide sequence. A 5' control region (*e.g.*, promoter) extends from nucleotides 1

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Before describing the present invention in detail, it is to be understood that this invention is not limited to particularly exemplified molecules or process parameters as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments of the invention only, and is not intended to be limiting. In addition, the practice of the present invention will employ, unless otherwise indicated, conventional methods of plant biology, virology, microbiology, molecular biology, recombinant DNA techniques and immunology all of which are within the ordinary skill of the art. Such techniques are explained fully in the literature. See, e.g., Evans, et al., *Handbook of Plant Cell Culture* (1983, Macmillan Publishing Co.); Binding, *Regeneration of Plants, Plant Protoplasts* (1985, CRC Press); Sambrook, et al., *Molecular Cloning: A Laboratory Manual* (2nd Edition, 1989); *DNA Cloning: A Practical Approach*, vol. I & II (D. Glover, ed.); *Oligonucleotide Synthesis* (N. Gait, ed., 1984); *A Practical Guide to Molecular Cloning* (1984); and *Fundamental Virology*, 2nd Edition, vol. I & II (B.N. Fields and D.M. Knipe, eds.).

All publications, patents and patent applications cited herein, whether *supra* or *infra*, are hereby incorporated by reference in their entirety.

It must be noted that, as used in this specification and the appended claims, the singular forms "a", "an" and "the" include plural referents unless the content clearly dictates otherwise. Thus, for example, reference to "a polypeptide" includes a mixture of two or more polypeptides, and the like.

The following amino acid abbreviations are used throughout the text:

	Alanine: Ala (A)	Arginine: Arg (R)
	Asparagine: Asn (N)	Aspartic acid: Asp (D)
25	Cysteine: Cys (C)	Glutamine: Gln (Q)
	Glutamic acid: Glu (E)	Glycine: Gly (G)
	Histidine: His (H)	Isoleucine: Ile (I)
	Leucine: Leu (L)	Lysine: Lys (K)
	Methionine: Met (M)	Phenylalanine: Phe (F)
30	Proline: Pro (P)	Serine: Ser (S)
	Threonine: Thr (T)	Tryptophan: Trp (W)
	Tyrosine: Tyr (Y)	Valine: Val (V)

Definitions

In describing the present invention, the following terms will be employed, and are intended to be defined as indicated below.

The terms "nucleic acid molecule" and "polynucleotide" are used interchangeably and refer to a polymeric form of nucleotides of any length, either deoxyribonucleotides or ribonucleotides, or analogs thereof. This term refers only to the primary structure of the molecule and thus includes double- and single-stranded DNA and RNA. It also includes known types of modifications, for example, labels which are known in the art, methylation, "caps", substitution of one or more of the naturally occurring nucleotides with an analog, internucleotide modifications such as, for example, those with uncharged linkages (e.g., methyl phosphonates, phosphotriesters, phosphoamidates, carbamates, etc.) and with charged linkages (e.g., phosphorothioates, phosphorodithioates, etc.), those containing pendant moieties, such as, for example proteins (including e.g., nucleases, toxins, antibodies, signal peptides, poly-L-lysine, etc.), those with intercalators (e.g., acridine, psoralen, etc.), those containing chelates (e.g., metals, radioactive metals, boron, oxidative metals, etc.), those containing alkylators, those with modified linkages (e.g., alpha anomeric nucleic acids, etc.), as well as unmodified forms of the polynucleotide.

Polynucleotides may have any three-dimensional structure, and may perform any function, known or unknown. Nonlimiting examples of polynucleotides include a gene, a gene fragment, exons, introns, messenger RNA (mRNA), transfer RNA, ribosomal RNA, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes, and primers.

A polynucleotide is typically composed of a specific sequence of four nucleotide bases: adenine (A); cytosine (C); guanine (G); and thymine (T) (uracil (U) for thymine (T) when the polynucleotide is RNA). Thus, the term polynucleotide sequence is the alphabetical representation of a polynucleotide molecule. This alphabetical representation can be input into databases in a computer having a central processing unit and used for bioinformatics applications such as functional genomics and homology searching.

Techniques for determining nucleic acid and amino acid “sequence identity” are known in the art. Typically, such techniques include determining the nucleotide sequence of the mRNA for a gene and/or determining the amino acid sequence encoded thereby, and comparing these sequences to a second nucleotide or amino acid sequence. In general, “identity” refers to an exact nucleotide-to-nucleotide or amino acid-to-amino acid correspondence of two polynucleotides or polypeptide sequences, respectively. Two or more sequences (polynucleotide or amino acid) can be compared by determining their “percent identity.” The percent identity of two sequences, whether nucleic acid or amino acid sequences, is the number of exact matches between two aligned sequences divided by the length of the shorter sequences and multiplied by 100. An approximate alignment for nucleic acid sequences is provided by the local homology algorithm of Smith and Waterman, Advances in Applied Mathematics 2:482-489 (1981). This algorithm can be applied to amino acid sequences by using the scoring matrix developed by Dayhoff, Atlas of Protein Sequences and Structure, M.O. Dayhoff ed., 5 suppl. 3:353-358, National Biomedical Research Foundation, Washington, D.C., USA, and normalized by Gribskov, Nucl. Acids Res. 14(6):6745-6763 (1986). An exemplary implementation of this algorithm to determine percent identity of a sequence is provided by the Genetics Computer Group (Madison, WI) in the “BestFit” utility application. The default parameters for this method are described in the Wisconsin Sequence Analysis Package Program Manual, Version 8 (1995) (available from Genetics Computer Group, Madison, WI). A preferred method of establishing percent identity in the context of the present invention is to use the MPSRCH package of programs copyrighted by the University of Edinburgh, developed by John F. Collins and Shane S. Sturrok, and distributed by IntelliGenetics, Inc. (Mountain View, CA). From this suite of packages the Smith-Waterman algorithm can be employed where default parameters are used for the scoring table (for example, gap open penalty of 12, gap extension penalty of one, and a gap of six). From the data generated the “Match” value reflects “sequence identity.” Other suitable programs for calculating the percent identity or similarity between sequences are generally known in the art, for example, another alignment program is BLAST, used with default parameters. For example, BLASTN and BLASTP can be used using the following default parameters: genetic

code = standard; filter = none; strand = both; cutoff = 60; expect = 10; Matrix = BLOSUM62; Descriptions = 50 sequences; sort by = HIGH SCORE; Databases = non-redundant, GenBank + EMBL + DDBJ + PDB + GenBank CDS translations + Swiss protein + Spupdate + PIR. Details of these programs can be found at the
5 following internet address: <http://www.ncbi.nlm.gov/cgi-bin/BLAST>.

Alternatively, the degree of sequence similarity between polynucleotides can be determined by hybridization of polynucleotides under conditions that form stable duplexes between homologous regions, followed by digestion with single-stranded-specific nuclease(s), and size determination of the digested fragments. Two DNA, or
10 two polypeptide sequences are "substantially homologous" to each other when the sequences exhibit at least about 43%-60%, preferably 60-70%, more preferably 70%-85%, more preferably at least about 85%-90%, more preferably at least about 90%-95%, and most preferably at least about 95%-98% sequence identity over a defined length of the molecules, or any percentage between the above-specified ranges, as
15 determined using the methods above. As used herein, substantially homologous also refers to sequences showing complete identity to the specified DNA or polypeptide sequence. DNA sequences that are substantially homologous can be identified in a Southern hybridization experiment under, for example, stringent conditions, as defined for that particular system. Defining appropriate hybridization conditions is
20 within the skill of the art. See, e.g., Sambrook et al., *supra*; *DNA Cloning, supra*; *Nucleic Acid Hybridization, supra*.

The degree of sequence identity between two nucleic acid molecules affects the efficiency and strength of hybridization events between such molecules. A partially identical nucleic acid sequence will at least partially inhibit a completely
25 identical sequence from hybridizing to a target molecule. Inhibition of hybridization of the completely identical sequence can be assessed using hybridization assays that are well known in the art (e.g., Southern blot, Northern blot, solution hybridization, or the like, see Sambrook, et al., *Molecular Cloning: A Laboratory Manual*, Second Edition, (1989) Cold Spring Harbor, N.Y.). Such assays can be conducted using
30 varying degrees of selectivity, for example, using conditions varying from low to high stringency. If conditions of low stringency are employed, the absence of non-specific binding can be assessed using a secondary probe that lacks even a partial degree of

sequence identity (for example, a probe having less than about 30% sequence identity with the target molecule), such that, in the absence of non-specific binding events, the secondary probe will not hybridize to the target.

When utilizing a hybridization-based detection system, a nucleic acid probe is
5 chosen that is complementary to a target nucleic acid sequence, and then by selection of appropriate conditions the probe and the target sequence “selectively hybridize,” or bind, to each other to form a hybrid molecule. A nucleic acid molecule that is capable of hybridizing selectively to a target sequence under “moderately stringent” typically hybridizes under conditions that allow detection of a target nucleic acid sequence of at
10 least about 10-14 nucleotides in length having at least approximately 70% sequence identity with the sequence of the selected nucleic acid probe. Stringent hybridization conditions typically allow detection of target nucleic acid sequences of at least about 10-14 nucleotides in length having a sequence identity of greater than about 90-95% with the sequence of the selected nucleic acid probe. Hybridization conditions useful
15 for probe/target hybridization where the probe and target have a specific degree of sequence identity, can be determined as is known in the art (see, for example, Nucleic Acid Hybridization: A Practical Approach, editors B.D. Hames and S.J. Higgins, (1985) Oxford; Washington, DC; IRL Press).

With respect to stringency conditions for hybridization, it is well known in the
20 art that numerous equivalent conditions can be employed to establish a particular stringency by varying, for example, the following factors: the length and nature of probe and target sequences, base composition of the various sequences, concentrations of salts and other hybridization solution components, the presence or absence of blocking agents in the hybridization solutions (e.g., formamide, dextran
25 sulfate, and polyethylene glycol), hybridization reaction temperature and time parameters, as well as, varying wash conditions. The selection of a particular set of hybridization conditions is selected following standard methods in the art (see, for example, Sambrook, et al., Molecular Cloning: A Laboratory Manual, Second Edition, (1989) Cold Spring Harbor, N.Y.).

30 A “gene” as used in the context of the present invention is a sequence of nucleotides in a genetic nucleic acid (chromosome, plasmid, etc.) with which a genetic function is associated. A gene is a hereditary unit, for example of an

organism, comprising a polynucleotide sequence that occupies a specific physical location (a "gene locus" or "genetic locus") within the genome of an organism. A gene can encode an expressed product, such as a polypeptide or a polynucleotide (e.g., tRNA). Alternatively, a gene may define a genomic location for a particular
5 event/function, such as the binding of proteins and/or nucleic acids, wherein the gene does not encode an expressed product. Typically, a gene includes coding sequences, such as, polypeptide encoding sequences, and non-coding sequences, such as, promoter sequences, polyadenylation sequences, transcriptional regulatory sequences (e.g., enhancer sequences). Many eucaryotic genes have "exons" (coding sequences)
10 interrupted by "introns" (non-coding sequences). In certain cases, a gene may share sequences with another gene(s) (e.g., overlapping genes).

A "coding sequence" or a sequence which "encodes" a selected polypeptide, is a nucleic acid molecule which is transcribed (in the case of DNA) and translated (in the case of mRNA) into a polypeptide, for example, *in vivo* when placed under the
15 control of appropriate regulatory sequences (or "control elements"). The boundaries of the coding sequence are typically determined by a start codon at the 5' (amino) terminus and a translation stop codon at the 3' (carboxy) terminus. A coding sequence can include, but is not limited to, cDNA from viral, procaryotic or eucaryotic mRNA, genomic DNA sequences from viral or procaryotic DNA, and even synthetic DNA
20 sequences. A transcription termination sequence may be located 3' to the coding sequence. Other "control elements" may also be associated with a coding sequence. A DNA sequence encoding a polypeptide can be optimized for expression in a selected cell by using the codons preferred by the selected cell to represent the DNA copy of the desired polypeptide coding sequence. "Encoded by" refers to a nucleic
25 acid sequence which codes for a polypeptide sequence, wherein the polypeptide sequence or a portion thereof contains an amino acid sequence of at least 3 to 5 amino acids, more preferably at least 8 to 10 amino acids, and even more preferably at least 15 to 20 amino acids from a polypeptide encoded by the nucleic acid sequence. Also encompassed are polypeptide sequences which are immunologically identifiable with
30 a polypeptide encoded by the sequence.

Typical "control elements", include, but are not limited to, transcription promoters, transcription enhancer elements, transcription termination signals,

polyadenylation sequences (located 3' to the translation stop codon), sequences for optimization of initiation of translation (located 5' to the coding sequence), translation enhancing sequences, and translation termination sequences. Transcription promoters can include inducible promoters (where expression of a polynucleotide sequence operably linked to the promoter is induced by an analyte, cofactor, regulatory protein, etc.), tissue-specific promoters (where expression of a polynucleotide sequence operably linked to the promoter is induced only in selected tissue), repressible promoters (where expression of a polynucleotide sequence operably linked to the promoter is induced by an analyte, cofactor, regulatory protein, etc.), and constitutive promoters.

A control element, such as a promoter, "directs the transcription" of a coding sequence in a cell when RNA polymerase will bind the promoter and transcribe the coding sequence into mRNA, which is then translated into the polypeptide encoded by the coding sequence.

"Expression enhancing sequences" typically refer to control elements that improve transcription or translation of a polynucleotide relative to the expression level in the absence of such control elements (for example, promoters, promoter enhancers, enhancer elements, and translational enhancers (e.g., Shine and Delagarno sequences)).

"Operably linked" refers to a juxtaposition wherein the components so described are in a relationship permitting them to function in their intended manner. A control sequence "operably linked" to a coding sequence is ligated in such a way that expression of the coding sequence is achieved under conditions compatible with the control sequences. The control elements need not be contiguous with the coding sequence, so long as they function to direct the expression thereof. Thus, for example, intervening untranslated yet transcribed sequences can be present between a promoter and the coding sequence and the promoter can still be considered "operably linked" to the coding sequence.

A "heterologous sequence" as used herein typically refers to a nucleic acid sequence that is not normally found in the cell or organism of interest. For example, a DNA sequence encoding a polypeptide can be obtained from a plant cell and

introduced into a bacterial cell. In this case the plant DNA sequence is "heterologous" to the native DNA of the bacterial cell.

The "native sequence" or "wild-type sequence" of a gene is the polynucleotide sequence that comprises the genetic locus corresponding to the gene, e.g., all
5 regulatory and open-reading frame coding sequences required for expression of a completely functional gene product as they are present in the wild-type genome of an organism. The native sequence of a gene can include, for example, transcriptional promoter sequences, translation enhancing sequences, introns, exons, and poly-A processing signal sites. It is noted that in the general population, wild-type genes may
10 include multiple prevalent versions that contain alterations in sequence relative to each other and yet do not cause a discernible pathological effect. These variations are designated "polymorphisms" or "allelic variations."

"Recombinant" as used herein to describe a nucleic acid molecule means a polynucleotide of genomic, cDNA, semisynthetic, or synthetic origin which, by virtue
15 of its origin or manipulation: (1) is not associated with all or a portion of the polynucleotide with which it is associated in nature; and/or (2) is linked to a polynucleotide other than that to which it is linked in nature. The term "recombinant" as used with respect to a protein or polypeptide means a polypeptide produced by expression of a recombinant polynucleotide.

20 By "vector" is meant any genetic element, such as a plasmid, phage, transposon, cosmid, chromosome, virus etc., which is capable of transferring gene sequences to target cells. Generally, a vector is capable of replication when associated with the proper control elements. Thus, the term includes cloning and expression vehicles, as well as viral vectors and integrating vectors.

25 As used herein, the term "expression cassette" refers to a molecule comprising at least one coding sequence operably linked to a control sequence which includes all nucleotide sequences required for the transcription of cloned copies of the coding sequence and the translation of the mRNAs in an appropriate host cell. Such expression cassettes can be used to express eukaryotic genes in a variety of hosts such
30 as bacteria, blue-green algae, plant cells, yeast cells, insect cells and animal cells. Under the invention, expression cassettes can include, but are not limited to, cloning vectors, specifically designed plasmids, viruses or virus particles. The cassettes may

further include an origin of replication for autonomous replication in host cells, selectable markers, various restriction sites, a potential for high copy number and strong promoters.

A cell has been "transformed" by an exogenous polynucleotide when the polynucleotide has been introduced inside the cell. The exogenous polynucleotide may or may not be integrated (covalently linked) into chromosomal DNA making up the genome of the cell. In prokaryotes and yeasts, for example, the exogenous DNA may be maintained on an episomal element, such as a plasmid. With respect to eucaryotic cells, a stably transformed cell is one in which the exogenous DNA has become integrated into the chromosome so that it is inherited by daughter cells through chromosome replication. This stability is demonstrated by the ability of the eucaryotic cell to establish cell lines or clones comprised of a population of daughter cells containing the exogenous DNA.

"Recombinant host cells," "host cells," "cells," "cell lines," "cell cultures," and other such terms denoting procaryotic microorganisms or eucaryotic cell lines cultured as unicellular entities, are used interchangeably, and refer to cells which can be, or have been, used as recipients for recombinant vectors or other transfer DNA, and include the progeny of the original cell which has been transfected. It is understood that the progeny of a single parental cell may not necessarily be completely identical in morphology or in genomic or total DNA complement to the original parent, due to accidental or deliberate mutation. Progeny of the parental cell which are sufficiently similar to the parent to be characterized by the relevant property, such as the presence of a nucleotide sequence encoding a desired peptide, are included in the progeny intended by this definition, and are covered by the above terms.

The term "*dwf4* polynucleotide" refers to a polynucleotide derived from the *dwf4* gene. The gene encodes the protein referred to herein as DWF4. DWF4 is a cytochrome P450 that mediates multiple 22 α -hydroxylation steps in brassinosteroid biosynthesis (see, Figure 1). The *dwf4* polynucleotide sequence and corresponding amino acid sequence are shown in Figures 10 and 11 (SEQ ID NO:1, SEQ ID NO:2 and GenBank accession No. AF044216). As shown in Figure 10, the *dwf4* coding sequence spans the region from nucleotide positions 3203 to 6110 and

the upstream 5' UTR, including the promoter region, spans nucleotide positions 1 to 3202. A functional 1.1 kb control element is also described in the Examples. A 3' UTR spans nucleotide positions 6111 to approximately 6468 of SEQ ID NO:1. The term as used herein encompasses a polynucleotide including a native sequence

5 depicted in Figure 10, as well as modifications and fragments thereof.

The term encompasses alterations to the polynucleotide sequence, so long as the alteration results in a plant displaying one or more *dwf4* phenotypic traits (described below) when the polynucleotide is expressed in a plant. Such modifications typically include deletions, additions and substitutions, to the native

10 *dwf4* sequence, so long as the mutation results in a plant displaying a *dwf4* phenotype as defined below. These modifications may be deliberate, as through site-directed mutagenesis, or may be accidental, such as through mutations of plants which express the *dwf4* polynucleotide or errors due to PCR amplification. The term encompasses expressed allelic variants of the wild-type *dwf4* sequence which may occur by normal

15 genetic variation or are produced by genetic engineering methods and which result in a detectable change in the wild-type *dwf4* phenotype.

The term "*dwf4* phenotype" as used herein refers to any microscopic or macroscopic change in structure or morphology of a plant, such as a transgenic plant, as well as biochemical differences, which are characteristic of a *dwf4* plant, compared

20 to a progenitor, wild-type plant cultivated under the same conditions. Generally, morphological differences include multiple short stems, short rounded leaves, loss of fertility due to reduced stamen length, and delayed development. Dark-grown *dwf4* seedlings possess short hypocotyls, open cotyledons, and developing leaves. The height of such plants will typically be 75% or less of the wild-type plant, more

25 typically 50% or less of the wild-type plant, and even more typically 25% or less of the wild-type plant, or any integer in between. Additional phenotypic morphological attributes of the *dwf4* mutant are summarized in Table 1 of the examples. Biochemically, *dwf4* hypocotyls are converted to wild-type length with the application of BL.

30 A "polypeptide" is used in it broadest sense to refer to a compound of two or more subunit amino acids, amino acid analogs, or other peptidomimetics. The subunits may be linked by peptide bonds or by other bonds, for example ester, ether,

etc. As used herein, the term "amino acid" refers to either natural and/or unnatural or synthetic amino acids, including glycine and both the D or L optical isomers, and amino acid analogs and peptidomimetics. A peptide of three or more amino acids is commonly called an oligopeptide if the peptide chain is short. If the peptide chain is long, the peptide is typically called a polypeptide or a protein. Full-length proteins, analogs, mutants and fragments thereof are encompassed by the definition. The terms also include postexpression modifications of the polypeptide, for example, glycosylation, acetylation, phosphorylation and the like. Furthermore, as ionizable amino and carboxyl groups are present in the molecule, a particular polypeptide may be obtained as an acidic or basic salt, or in neutral form. A polypeptide may be obtained directly from the source organism, or may be recombinantly or synthetically produced (see further below).

A "DWF4" polypeptide is a polypeptide as defined above, which is derived from a 22 α -hydroxylase that functions in the brassinolide (BL) biosynthetic pathway (see, Figure 1). The native sequence of full-length DWF4 is shown in Figure 11 (SEQ ID NO:2). However, the term encompasses mutants and fragments of the native sequence so long as the protein functions for its intended purpose.

The term "DWF4 analog" refers to derivatives of DWF4, or fragments of such derivatives, that retain desired function, e.g., as measured in assays as described further below. In general, the term "analog" refers to compounds having a native polypeptide sequence and structure with one or more amino acid additions, substitutions (generally conservative in nature) and/or deletions, relative to the native molecule, so long as the modifications do not destroy desired activity. Preferably, the analog has at least the same activity as the native molecule. Methods for making polypeptide analogs are known in the art and are described further below.

Particularly preferred analogs include substitutions that are conservative in nature, i.e., those substitutions that take place within a family of amino acids that are related in their side chains. Specifically, amino acids are generally divided into four families: (1) acidic -- aspartate and glutamate; (2) basic -- lysine, arginine, histidine; (3) non-polar -- alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan; and (4) uncharged polar -- glycine, asparagine, glutamine, cysteine, serine threonine, tyrosine. Phenylalanine, tryptophan, and tyrosine are

sometimes classified as aromatic amino acids. For example, it is reasonably predictable that an isolated replacement of leucine with isoleucine or valine, an aspartate with a glutamate, a threonine with a serine, or a similar conservative replacement of an amino acid with a structurally related amino acid, will not have a major effect on the biological activity. It is to be understood that the terms include the various sequence polymorphisms that exist, wherein amino acid substitutions in the protein sequence do not affect the essential functions of the protein.

By "purified" and "isolated" is meant, when referring to a polypeptide or polynucleotide, that the molecule is separate and discrete from the whole organism with which the molecule is found in nature; or devoid, in whole or part, of sequences normally associated with it in nature; or a sequence, as it exists in nature, but having heterologous sequences (as defined below) in association therewith. It is to be understood that the term "isolated" with reference to a polynucleotide intends that the polynucleotide is separate and discrete from the chromosome from which the polynucleotide may derive. The term "purified" as used herein preferably means at least 75% by weight, more preferably at least 85% by weight, more preferably still at least 95% by weight, and most preferably at least 98% by weight, of biological macromolecules of the same type are present. An "isolated polynucleotide which encodes a particular polypeptide" refers to a nucleic acid molecule which is substantially free of other nucleic acid molecules that do not encode the subject polypeptide; however, the molecule may include some additional bases or moieties which do not deleteriously affect the basic characteristics of the composition.

By "fragment" is intended a polypeptide or polynucleotide consisting of only a part of the intact sequence and structure of the reference polypeptide or polynucleotide, respectively. The fragment can include a 3' or C-terminal deletion or a 5' or N-terminal deletion, or even an internal deletion, of the native molecule. A polynucleotide fragment of a *dwf4* sequence will generally include at least about 15 contiguous bases of the molecule in question, more preferably 18-25 contiguous bases, even more preferably 30-50 or more contiguous bases of the *dwf4* molecule, or any integer between 15 bases and the full-length sequence of the molecule. Fragments which provide at least one *dwf4* phenotype as defined above are useful in

the production of transgenic plants. Fragments are also useful as oligonucleotide probes, to find additional *dwf4* sequences.

Similarly, a polypeptide fragment of a DWF4 molecule will generally include at least about 5-10 contiguous amino acid residues of the full-length molecule, preferably at least about 15-25 contiguous amino acid residues of the full-length molecule, and most preferably at least about 20-50 or more contiguous amino acid residues of the full-length DWF4 molecule, or any integer between 10 amino acids and the full-length sequence of the molecule. Such fragments are useful for the production of antibodies and the like.

By "transgenic plant" is meant a plant into which one or more exogenous polynucleotides have been introduced. Examples of means by which this can be accomplished are described below, and include *Agrobacterium*-mediated transformation, biolistic methods, electroporation, and the like. In the context of the present invention, the transgenic plant contains a polynucleotide which is not normally present in the corresponding wild-type plant and which confers at least one *dwf4* phenotypic trait to the plant. The transgenic plant therefore exhibits altered structure, morphology or biochemistry as compared with a progenitor plant which does not contain the transgene, when the transgenic plant and the progenitor plant are cultivated under similar or equivalent growth conditions. Such a plant containing the exogenous polynucleotide is referred to here as an R_1 generation transgenic plant. Transgenic plants may also arise from sexual cross or by selfing of transgenic plants into which exogenous polynucleotides have been introduced. Such a plant containing the exogenous nucleic acid is also referred to here as an R_1 generation transgenic plant. Transgenic plants which arise from a sexual cross with another parent line or by selfing are "descendants or the progeny" of a R_1 plant and are generally called F_n plants or S_n plants, respectively, n meaning the number of generations.

General Overview

In this report, we present morphological, biochemical, and molecular analysis of a novel gene, *dwf4*, isolated from *Arabidopsis*. Morphologically, *dwf4* plants display a dramatic reduction in the length of many different organs examined, and this size reduction is attributable to a defect in cell elongation. Biochemically, *dwf4*

hypocotyls were converted completely to wild-type length with the application of BL, suggesting a deficiency in BRs. In agreement with this, BR intermediate feeding analysis, indicated that *dwf4* encodes a cytochrome P450 that mediates multiple 22 α -hydroxylation steps in brassinosteroid biosynthesis. Sequencing of the *dwf4* locus and analysis of the protein product are described.

The molecules of the present invention are therefore useful in the production of transgenic plants which display at least one *dwf4* phenotype, so that the resulting plants have altered structure or morphology. The present invention particularly provides for altered structure or morphology such as reduced cell length, extended flowering periods, increased size of leaves or fruit, increased branching, increased seed production and altered sterol composition relative wild-type plants. The DWF4 polypeptides can be expressed to engineer a plant with desirable properties. The engineering is accomplished by transforming plants with nucleic acid constructs described herein which may also comprise promoters and secretion signal peptides. The transformed plants or their progenies are screened for plants that express the desired polypeptide.

Engineered plants exhibiting the desired altered structure or morphology can be used in plant breeding or directly in agricultural production or industrial applications. Plants having the altered polypeptide can be crossed with other altered plants engineered with alterations in other growth modulation enzymes, proteins or polypeptides to produce lines with even further enhanced altered structural morphology characteristics compared to the parents or progenitor plants.

Isolation of Nucleic Acid Sequences from Plants

The isolation of *dwf4* sequences from the polynucleotides of the invention may be accomplished by a number of techniques. For instance, oligonucleotide probes based on the sequences disclosed here can be used to identify the desired gene in a cDNA or genomic DNA library from a desired plant species. To construct genomic libraries, large segments of genomic DNA are generated by random fragmentation, e.g. using restriction endonucleases, and are ligated with vector DNA to form concatemers that can be packaged into the appropriate vector. To prepare a

library of tissue-specific cDNAs, mRNA is isolated from tissues and a cDNA library which contains the gene transcripts is prepared from the mRNA.

The cDNA or genomic library can then be screened using a probe based upon the sequence of a cloned gene such as the polynucleotides disclosed here. Probes may
5 be used to hybridize with genomic DNA or cDNA sequences to isolate homologous genes in the same or different plant species. Alternatively, the nucleic acids of interest can be amplified from nucleic acid samples using amplification techniques. For instance, polymerase chain reaction (PCR) technology to amplify the sequences of the genes directly from mRNA, from cDNA, from genomic libraries or cDNA
10 libraries. PCR-RTM. and other *in vitro* amplification methods may also be useful, for example, to clone nucleic acid sequences that code for proteins to be expressed, to make nucleic acids to use as probes for detecting the presence of the desired mRNA in samples, for nucleic acid sequencing, or for other purposes.

Appropriate primers and probes for identifying *dwf4*-specific genes from plant
15 tissues are generated from comparisons of the sequences provided herein. For a general overview of PCR see Innis et al. eds, *PCR Protocols: A Guide to Methods and Applications*, Academic Press, San Diego (1990). Appropriate primers for this invention include, for instance, those primers described in the Examples and Sequence Listings, as well as other primers derived from the *dwf4* sequences disclosed herein.
20 Suitable amplifications conditions may be readily determined by one of skill in the art in view of the teachings herein, for example, including reaction components and amplification conditions as follows: 10 mM Tris-HCl, pH 8.3, 50 mM potassium chloride, 1.5 mM magnesium chloride, 0.001% gelatin, 200 μ M dATP, 200 μ M dCTP, 200 μ M dGTP, 200 μ M dTTP, 0.4 μ M primers, and 100 units per mL Taq
25 polymerase; 96°C for 3 min., 30 cycles of 96°C for 45 seconds, 50°C for 60 seconds, 72°C for 60 seconds, followed by 72°C for 5 min.

Polynucleotides may also be synthesized by well-known techniques as described in the technical literature. See, e.g., Carruthers, et al. (1982) *Cold Spring Harbor Symp. Quant. Biol.* 47:411-418, and Adams, et al. (1983) *J. Am. Chem. Soc.*
30 105:661. Double stranded DNA fragments may then be obtained either by synthesizing the complementary strand and annealing the strands together under

appropriate conditions, or by adding the complementary strand using DNA polymerase with an appropriate primer sequence.

The polynucleotides of the present invention may also be used to isolate or create other mutant cell gene alleles. Mutagenesis consists primarily of site-directed
5 mutagenesis followed by phenotypic testing of the altered gene product. Some of the more commonly employed site-directed mutagenesis protocols take advantage of vectors that can provide single stranded as well as double stranded DNA, as needed. Generally, the mutagenesis protocol with such vectors is as follows. A mutagenic primer, i.e., a primer complementary to the sequence to be changed, but consisting of
10 one or a small number of altered, added, or deleted bases, is synthesized. The primer is extended *in vitro* by a DNA polymerase and, after some additional manipulations, the now double-stranded DNA is transfected into bacterial cells. Next, by a variety of methods, the desired mutated DNA is identified, and the desired protein is purified from clones containing the mutated sequence. For longer sequences, additional
15 cloning steps are often required because long inserts (longer than 2 kilobases) are unstable in those vectors. Protocols are known to one skilled in the art and kits for site-directed mutagenesis are widely available from biotechnology supply companies, for example from Amersham Life Science, Inc. (Arlington Heights, Ill.) and Stratagene Cloning Systems (La Jolla, Calif.).

20

Control elements

Regulatory regions can be isolated from the *dwf4* gene and used in recombinant constructs for modulating the expression of the *dwf4* gene or a heterologous gene *in vitro* and/or *in vivo*. As shown in Figure 10, the coding region
25 of the *dwf4* gene (designated by the light grey bar) begins at nucleotide position 1133. The region of the gene spanning nucleotide positions 990-1132 of Figure 10 includes the *dwf4* promoter. This region may be used in its entirety or fragments of the region may be isolated which provide the ability to direct expression of a coding sequence linked thereto.

30

Thus, promoters can be identified by analyzing the 5' sequences of a genomic clone corresponding to the *dwf4*-specific genes described here. Sequences characteristic of promoter sequences can be used to identify the promoter. Sequences

controlling eukaryotic gene expression have been extensively studied. For instance, promoter sequence elements include the TATA box consensus sequence (TATAAT), which is usually 20 to 30 base pairs upstream of the transcription start site. In most instances the TATA box is required for accurate transcription initiation. In plants,
5 further upstream from the TATA box, at positions -80 to -100, there is typically a promoter element with a series of adenines surrounding the trinucleotide G (or T) N G. (See, J. Messing et al., in *Genetic Engineering in Plants*, pp. 221-227 (Kosage, Meredith and Hollaender, eds. (1983)). Methods for identifying and characterizing promoter regions in plant genomic DNA are described, for example, in Jordano et al.
10 (1989) *Plant Cell* 1:855-866; Bustos et al (1989) *Plant Cell* 1:839-854; Green et al. (1988) *EMBO J.* 7:4035-4044; Meier et al. (1991) *Plant Cell* 3:309-316; and Zhang et al (1996) *Plant Physiology* 110:1069-1079).

Additionally, the promoter region may include nucleotide substitutions, insertions or deletions that do not substantially affect the binding of relevant DNA
15 binding proteins and hence the promoter function. It may, at times, be desirable to decrease the binding of relevant DNA binding proteins to "silence" or "down-regulate" a promoter, or conversely to increase the binding of relevant DNA binding proteins to "enhance" or "up-regulate" a promoter. In such instances, the nucleotide sequence of the promoter region may be modified by, e.g., inserting additional
20 nucleotides, changing the identity of relevant nucleotides, including use of chemically-modified bases, or by deleting one or more nucleotides.

Promoter function can be assayed by methods known in the art, preferably by measuring activity of a reporter gene operatively linked to the sequence being tested for promoter function. Examples of reporter genes include those encoding luciferase,
25 green fluorescent protein, GUS, neo, cat and bar.

Polynucleotides comprising untranslated (UTR) sequences and intron/exon junctions are also within the scope of the invention. UTR sequences include introns and 5' or 3' untranslated regions (5' UTRs or 3' UTRs). As shown in Figures 2 and 10, the *dwf4* gene sequence includes eight exons and seven introns. These portions of
30 the *dwf4* gene especially UTRs, can have regulatory functions related to, for example, translation rate and mRNA stability. Thus, these portions of the gene can be isolated for use as elements of gene constructs for expression of polynucleotides encoding

desired polypeptides. The 5' control element region of *dwf4* extends from nucleotides 1 through 3202 of SEQ ID NO:1. Further, as described in Example 11, a 1.1 kb portion of this region that is directly upstream of the translation initiation site contains elements necessary for transcriptional control of *dwf4*. In contrast, a 280 bp fragment of the *dwf4* control element region that includes the TATA-like region does not appear to contain all of the necessary transcriptional control elements (see, Example 11).

Introns of genomic DNA segments may also have regulatory functions. Sometimes promoter elements, especially transcription enhancer or suppressor elements, are found within introns. Also, elements related to stability of heteronuclear RNA and efficiency of transport to the cytoplasm for translation can be found in intron elements. Thus, these segments can also find use as elements of expression vectors intended for use to transform plants.

The introns, UTR sequences and intron/exon junctions can vary from the native sequence. Such changes from those sequences preferably will not affect the regulatory activity of the UTRs or intron or intron/exon junction sequences on expression, transcription, or translation. However, in some instances, down-regulation of such activity may be desired to modulate traits or phenotypic or *in vitro* activity.

20

Use of Nucleic Acids of the Invention to Inhibit Gene Expression

The isolated sequences prepared as described herein, can be used to prepare expression cassettes useful in a number of techniques. For example, expression cassettes of the invention can be used to suppress (underexpress) endogenous *dwf4* gene expression. Inhibiting expression can be useful, for instance, in suppressing the phenotype (*e.g.*, dwarf appearance, 22 α -hydroxylase activity) exhibited by *dwf4* plants. Further, the inhibitory polynucleotides of the present invention can also be used in combination with overexpressing constructs described below, for example, using suitable tissue-specific promoters linked to polynucleotides described herein. In this way, the polynucleotides can be used to promote *dwf4* phenotypes (*e.g.*, activity) in selected tissue and, at the same time, inhibit *dwf4* phenotypes (*e.g.*, activity) in different tissue(s).

30

A number of methods can be used to inhibit gene expression in plants. For instance, antisense technology can be conveniently used. To accomplish this, a nucleic acid segment from the desired gene is cloned and operably linked to a promoter such that the antisense strand of RNA will be transcribed. The expression
5 cassette is then transformed into plants and the antisense strand of RNA is produced. In plant cells, it has been suggested that antisense RNA inhibits gene expression by preventing the accumulation of mRNA which encodes the enzyme of interest, see, e.g., Sheehy et al (1988) *Proc. Nat. Acad. Sci. USA* 85:8805-8809, and Hiatt et al., U.S. Patent Number 4,801,340.

10 The nucleic acid segment to be introduced generally will be substantially identical to at least a portion of the endogenous gene or genes to be repressed. The sequence, however, need not be perfectly identical to inhibit expression. The vectors of the present invention can be designed such that the inhibitory effect applies to other proteins within a family of genes exhibiting homology or substantial homology to the
15 target gene.

For antisense suppression, the introduced sequence also need not be full length relative to either the primary transcription product or fully processed mRNA. Generally, higher homology can be used to compensate for the use of a shorter sequence. Furthermore, the introduced sequence need not have the same intron or
20 exon pattern, and homology of non-coding segments may be equally effective. Normally, a sequence of between about 30 or 40 nucleotides and about full length nucleotides should be used, though a sequence of at least about 100 nucleotides is preferred, a sequence of at least about 200 nucleotides is more preferred, and a sequence of at least about 500 nucleotides is especially preferred. It is to be
25 understood that any integer between the above-recited ranges is intended to be captured herein.

Catalytic RNA molecules or ribozymes can also be used to inhibit expression of *dwf4* genes. It is possible to design ribozymes that specifically pair with virtually any target RNA and cleave the phosphodiester backbone at a specific location,
30 thereby functionally inactivating the target RNA. In carrying out this cleavage, the ribozyme is not itself altered, and is thus capable of recycling and cleaving other molecules, making it a true enzyme. The inclusion of ribozyme sequences within

antisense RNAs confers RNA-cleaving activity upon them, thereby increasing the activity of the constructs.

A number of classes of ribozymes have been identified. One class of ribozymes is derived from a number of small circular RNAs which are capable of self-cleavage and replication in plants. The RNAs replicate either alone (viroid
5 RNAs) or with a helper virus (satellite RNAs). Examples include RNAs from avocado sunblotch viroid and the satellite RNAs from tobacco ringspot virus, lucerne transient streak virus, velvet tobacco mottle virus, solanum nodiflorum mottle virus and subterranean clover mottle virus. The design and use of target RNA-specific
10 ribozymes is described in Haseloff et al (1988) *Nature* 334:585-591.

Another method of suppression is sense suppression. Introduction of expression cassettes in which a nucleic acid is configured in the sense orientation with respect to the promoter has been shown to be an effective means by which to block the transcription of target genes. For an example of the use of this method to
15 modulate expression of endogenous genes see, Napoli et al (1990) *The Plant Cell* 2:279-289 and U.S. Patent Numbers 5,034,323, 5,231,020, and 5,283,184.

Generally, where inhibition of expression is desired, some transcription of the introduced sequence occurs. The effect may occur where the introduced sequence contains no coding sequence per se, but only intron or untranslated sequences
20 homologous to sequences present in the primary transcript of the endogenous sequence. The introduced sequence generally will be substantially identical to the endogenous sequence intended to be repressed. This minimal identity will typically be greater than about 50%-65%, but a higher identity might exert a more effective repression of expression of the endogenous sequences. Substantially greater identity
25 of more than about 80% is preferred, though about 95% to absolute identity would be most preferred. It is to be understood that any integer between the above-recited ranges is intended to be captured herein. As with antisense regulation, the effect should apply to any other proteins within a similar family of genes exhibiting homology or substantial homology.

30 For sense suppression, the introduced sequence in the expression cassette, needing less than absolute identity, also need not be full length, relative to either the primary transcription product or fully processed mRNA. This may be preferred to

avoid concurrent production of some plants which are overexpressers. A higher identity in a shorter than full length sequence compensates for a longer, less identical sequence. Furthermore, the introduced sequence need not have the same intron or exon pattern, and identity of non-coding segments will be equally effective.

- 5 Normally, a sequence of the size ranges noted above for antisense regulation is used.

Use of Nucleic Acids of the Invention to Enhance Gene Expression

- In addition to inhibiting certain features of a plant, the polynucleotides of the invention can be used to increase certain features such as extending flowering,
10 producing larger leaves or fruit, producing increased branching and increasing seed production. This can be accomplished by the overexpression of *dwf4* polynucleotides.

- The exogenous *dwf4* polynucleotides do not have to code for exact copies of the endogenous *dwf4* proteins. Modified DWF4 protein chains can also be readily designed utilizing various recombinant DNA techniques well known to those skilled
15 in the art and described for instance, in Sambrook et al., *supra*. Hydroxylamine can also be used to introduce single base mutations into the coding region of the gene (Sikorski et al (1991) *Meth. Enzymol.* 194: 302-318). For example, the chains can vary from the naturally occurring sequence at the primary structure level by amino acid substitutions, additions, deletions, and the like. These modifications can be used
20 in a number of combinations to produce the final modified protein chain.

- It will be apparent that the polynucleotides described herein can be used in a variety of combinations. For example, the polynucleotides can be used to produce different phenotypes in the same organism, for instance by using tissue-specific promoters to overexpress a *dwf4* polynucleotide in certain tissues (*e.g.*, leaf tissue)
25 while at the same time using tissue-specific promoters to inhibit expression of *dwf4* in other tissues. In addition, fusion proteins of the polynucleotides described herein with other known polynucleotides (*e.g.*, polynucleotides encoding products involved in the BR pathway) can be constructed and employed to obtain desired phenotypes.

- Any of the *dwf4* polynucleotides described herein can also be used in standard
30 diagnostic assays, for example, in assays mRNA levels (see, Sambrook et al, *supra*); as hybridization probes, *e.g.*, in combination with appropriate means, such as a label,

for detecting hybridization (see, Sambrook et al., *supra*); as primers, *e.g.*, for PCR (see, Sambrook et al., *supra*); attached to solid phase supports and the like.

Preparation of Recombinant Vectors

5 To use isolated sequences in the above techniques, recombinant DNA vectors suitable for transformation of plant cells are prepared. Techniques for transforming a wide variety of higher plant species are well known and described further below as well as in the technical and scientific literature. See, for example, Weising et al (1988) *Ann. Rev. Genet.* 22:421-477. A DNA sequence coding for the desired polypeptide,
10 for example a cDNA sequence encoding the full length DWF4 protein, will preferably be combined with transcriptional and translational initiation regulatory sequences which will direct the transcription of the sequence from the gene in the intended tissues of the transgenic plant.

 Such regulatory elements include but are not limited to the promoters derived
15 from the genome of plant cells (*e.g.*, heat shock promoters such as soybean hsp17.5-E or hsp17.3-B (Gurley et al. (1986) *Mol. Cell. Biol.* 6:559-565); the promoter for the small subunit of RUBISCO (Coruzzi et al. (1984) *EMBO J.* 3:1671-1680; Broglie et al (1984) *Science* 224:838-843); the promoter for the chlorophyll a/b binding protein) or from plant viruses viral promoters such as the 35S RNA and 19S RNA promoters
20 of CaMV (Brisson et al. (1984) *Nature* 310:511-514), or the coat protein promoter of TMV (Takamatsu et al. (1987) *EMBO J.* 6:307-311), cytomegalovirus hCMV immediate early gene, the early or late promoters of SV40 adenovirus, the lac system, the trp system, the TAC system, the TRC system, the major operator and promoter regions of phage A, the control regions of fd coat protein, the promoter for
25 3-phosphoglycerate kinase, the promoters of acid phosphatase, heat shock promoters (*e.g.*, as described above) and the promoters of the yeast alpha-mating factors.

 In construction of recombinant expression cassettes of the invention, a plant promoter fragment may be employed which will direct expression of the gene in all tissues of a regenerated plant. Such promoters are referred to herein as "constitutive"
30 promoters and are active under most environmental conditions and states of development or cell differentiation. Examples of constitutive promoters include the cauliflower mosaic virus (CaMV) 35S transcription initiation region, the T-DNA

mannopine synthetase promoter (e.g., the 1'- or 2'- promoter derived from T-DNA of *Agrobacterium tumefaciens*), and other transcription initiation regions from various plant genes known to those of skill.

Alternatively, the plant promoter may direct expression of the polynucleotide
5 of the invention in a specific tissue (tissue-specific promoters) or may be otherwise under more precise environmental control (inducible promoters). Examples of tissue-specific promoters under developmental control include promoters that initiate transcription only in certain tissues, such as fruit, seeds, or flowers such as tissue- or developmental-specific promoter, such as, but not limited to the *dwf4* promoter, the
10 CHS promoter, the PATATIN promoter, etc. The tissue specific E8 promoter from tomato is particularly useful for directing gene expression so that a desired gene product is located in fruits.

Other suitable promoters include those from genes encoding embryonic storage proteins. Examples of environmental conditions that may affect transcription
15 by inducible promoters include anaerobic conditions, elevated temperature, or the presence of light. If proper polypeptide expression is desired, a polyadenylation region at the 3'-end of the coding region should be included. The polyadenylation region can be derived from the natural gene, from a variety of other plant genes, or from T-DNA. In addition, the promoter itself can be derived from the *dwf4* gene, as
20 described above.

The vector comprising the sequences (e.g., promoters or coding regions) from genes of the invention will typically comprise a marker gene which confers a selectable phenotype on plant cells. For example, the marker may encode biocide resistance, particularly antibiotic resistance, such as resistance to kanamycin, G418,
25 bleomycin, hygromycin, or herbicide resistance, such as resistance to chlorosulfuron or Basta.

Production of Transgenic Plants

DNA constructs of the invention may be introduced into the genome of the
30 desired plant host by a variety of conventional techniques. For reviews of such techniques see, for example, Weissbach & Weissbach *Methods for Plant Molecular Biology* (1988, Academic Press, N.Y.) Section VIII, pp. 421-463; and Grierson &

Corey, *Plant Molecular Biology* (1988, 2d Ed.), Blackie, London, Ch. 7-9. For example, the DNA construct may be introduced directly into the genomic DNA of the plant cell using techniques such as electroporation and microinjection of plant cell protoplasts, or the DNA constructs can be introduced directly to plant tissue using
5 biolistic methods, such as DNA particle bombardment (see, e.g., Klein et al (1987) *Nature* 327:70-73). Alternatively, the DNA constructs may be combined with suitable T-DNA flanking regions and introduced into a conventional *Agrobacterium tumefaciens* host vector. *Agrobacterium tumefaciens*-mediated transformation techniques, including disarming and use of binary vectors, are well described in the
10 scientific literature. See, for example Horsch et al (1984) *Science* 233:496-498, and Fraley et al (1983) *Proc. Nat'l. Acad. Sci. USA* 80:4803. The virulence functions of the *Agrobacterium tumefaciens* host will direct the insertion of the construct and adjacent marker into the plant cell DNA when the cell is infected by the bacteria using binary T DNA vector (Bevan (1984) *Nuc. Acid Res.* 12:8711-8721) or the
15 co-cultivation procedure (Horsch et al (1985) *Science* 227:1229-1231). Generally, the *Agrobacterium* transformation system is used to engineer dicotyledonous plants (Bevan et al (1982) *Ann. Rev. Genet* 16:357-384; Rogers et al (1986) *Methods Enzymol.* 118:627-641). The *Agrobacterium* transformation system may also be used to transform, as well as transfer, DNA to monocotyledonous plants and plant cells.
20 (see Hernalsteen et al (1984) *EMBO J* 3:3039-3041; Hooykass-Van Slogteren et al (1984) *Nature* 311:763-764; Grimsley et al (1987) *Nature* 325:1677-179; Boulton et al (1989) *Plant Mol. Biol.* 12:31-40.; and Gould et al (1991) *Plant Physiol.* 95:426-434).

Alternative gene transfer and transformation methods include, but are not
25 limited to, protoplast transformation through calcium-, polyethylene glycol (PEG)- or electroporation-mediated uptake of naked DNA (see Paszkowski et al. (1984) *EMBO J* 3:2717-2722, Potrykus et al. (1985) *Molec. Gen. Genet.* 199:169-177; Fromm et al. (1985) *Proc. Nat. Acad. Sci. USA* 82:5824-5828; and Shimamoto (1989) *Nature* 338:274-276) and electroporation of plant tissues (D'Halluin et al. (1992) *Plant Cell*
30 4:1495-1505). Additional methods for plant cell transformation include microinjection, silicon carbide mediated DNA uptake (Kaepler et al. (1990) *Plant Cell Reporter* 9:415-418), and microprojectile bombardment (see Klein et al. (1988)

Proc. Nat. Acad. Sci. USA 85:4305-4309; and Gordon-Kamm et al. (1990) *Plant Cell* 2:603-618).

Transformed plant cells which are produced by any of the above transformation techniques can be cultured to regenerate a whole plant which possesses the transformed genotype and thus the desired phenotype. Such regeneration techniques rely on manipulation of certain phytohormones in a tissue culture growth medium, typically relying on a biocide and/or herbicide marker which has been introduced together with the desired nucleotide sequences. Plant regeneration from cultured protoplasts is described in Evans, et al., "Protoplasts Isolation and Culture" in *Handbook of Plant Cell Culture*, pp. 124-176, Macmillian Publishing Company, New York, 1983; and Binding, *Regeneration of Plants, Plant Protoplasts*, pp. 21-73, CRC Press, Boca Raton, 1985. Regeneration can also be obtained from plant callus, explants, organs, pollens, embryos or parts thereof. Such regeneration techniques are described generally in Klee et al (1987) *Ann. Rev. of Plant Phys.* 38:467-486.

The nucleic acids of the invention can be used to confer desired traits on essentially any plant. A wide variety of plants and plant cell systems may be engineered for the desired physiological and agronomic characteristics described herein using the nucleic acid constructs of the present invention and the various transformation methods mentioned above. In preferred embodiments, target plants and plant cells for engineering include, but are not limited to, those monocotyledonous and dicotyledonous plants, such as crops including grain crops (e.g., wheat, maize, rice, millet, barley), fruit crops (e.g., tomato, apple, pear, strawberry, orange), forage crops (e.g., alfalfa), root vegetable crops (e.g., carrot, potato, sugar beets, yam), leafy vegetable crops (e.g., lettuce, spinach); flowering plants (e.g., petunia, rose, chrysanthemum), conifers and pine trees (e.g., pine fir, spruce); plants used in phytoremediation (e.g., heavy metal accumulating plants); oil crops (e.g., sunflower, rape seed) and plants used for experimental purposes (e.g., Arabidopsis). Thus, the invention has use over a broad range of plants, including, but not limited to, species from the genera Asparagus, Avena, Brassica, Citrus, Citrullus, Capsicum, Cucurbita, Daucus, Glycine, Hordeum, Lactuca, Lycopersicon, Malus, Manihot, Nicotiana, Oryza, Persea, Pisum, Pyrus, Prunus, Raphanus, Secale, Solanum, Sorghum, Triticum, Vitis, Vigna, and Zea.

One of skill in the art will recognize that after the expression cassette is stably incorporated in transgenic plants and confirmed to be operable, it can be introduced into other plants by sexual crossing. Any of a number of standard breeding techniques can be used, depending upon the species to be crossed.

5 A transformed plant cell, callus, tissue or plant may be identified and isolated by selecting or screening the engineered plant material for traits encoded by the marker genes present on the transforming DNA. For instance, selection may be performed by growing the engineered plant material on media containing an inhibitory amount of the antibiotic or herbicide to which the transforming gene
10 construct confers resistance. Further, transformed plants and plant cells may also be identified by screening for the activities of any visible marker genes (e.g., the β -glucuronidase, luciferase, B or C1 genes) that may be present on the recombinant nucleic acid constructs of the present invention. Such selection and screening methodologies are well known to those skilled in the art.

15 Physical and biochemical methods also may be used to identify plant or plant cell transformants containing the gene constructs of the present invention. These methods include but are not limited to: 1) Southern analysis or PCR amplification for detecting and determining the structure of the recombinant DNA insert; 2) Northern blot, S1 RNase protection, primer-extension or reverse transcriptase-PCR
20 amplification for detecting and examining RNA transcripts of the gene constructs; 3) enzymatic assays for detecting enzyme or ribozyme activity, where such gene products are encoded by the gene construct; 4) protein gel electrophoresis, Western blot techniques, immunoprecipitation, or enzyme-linked immunoassays, where the gene construct products are proteins. Additional techniques, such as *in situ*
25 hybridization, enzyme staining, and immunostaining, also may be used to detect the presence or expression of the recombinant construct in specific plant organs and tissues. The methods for doing all these assays are well known to those skilled in the art.

 Effects of gene manipulation using the methods of this invention can be
30 observed by, for example, northern blots of the RNA (e.g., mRNA) isolated from the tissues of interest. Typically, if the amount of mRNA has increased, it can be assumed that the endogenous *dwf4* gene is being expressed at a greater rate than

before. Other methods of measuring DWF4 activity can be used. For example, cell length can be measured at specific times. Because *dwf4* affects the BR biosynthetic pathway, an assay that measures the amount of BL can also be used. Such assays are known in the art. Different types of enzymatic assays can be used, depending on the
5 substrate used and the method of detecting the increase or decrease of a reaction product or by-product. In addition, the levels of DWF4 protein expressed can be measured immunochemically, i.e., ELISA, RIA, EIA and other antibody based assays well known to those of skill in the art, by electrophoretic detection assays (either with staining or western blotting), and sterol (BL) detection assays.

10 The transgene may be selectively expressed in some tissues of the plant or at some developmental stages, or the transgene may be expressed in substantially all plant tissues, substantially along its entire life cycle. However, any combinatorial expression mode is also applicable.

The present invention also encompasses seeds of the transgenic plants
15 described above wherein the seed has the transgene or gene construct. The present invention further encompasses the progeny, clones, cell lines or cells of the transgenic plants described above wherein said progeny, clone, cell line or cell has the transgene or gene construct.

20 Polypeptides

The present invention also includes DWF4 polypeptides, including such polypeptides as a fusion, or chimeric protein product (comprising the protein, fragment, analogue, mutant or derivative joined via a peptide bond to a heterologous protein sequence (of a different protein)). Such a chimeric product can be made by
25 ligating the appropriate nucleic acid sequences encoding the desired amino acid sequences to each other by methods known in the art, in the proper coding frame, and expressing the chimeric product by methods commonly known in the art.

As noted above, DWF4 phenotype includes any macroscopic, microscopic or biochemical changes which are characteristic of over- or under-expression of *dwf4*.
30 Thus, DWF4 polypeptide phenotype (e.g., activities) can include any activity that is exhibited by the native DWF4 polypeptide including, for example, *in vitro*, *in vivo*,

biological, enzymatic, immunological, substrate binding activities, etc. Non-limiting examples of DWF4 activities include:

- (a) activities displayed by other heme-thiolate enzymes;
- (b) characteristic Soret absorption peak at 40 nm when the substrate-bound
5 reduced form is exposed to the lights (*see, e.g.,* Jefcoate et al., *infra*);
- (c) hydroxylation of various substrates via monooxygenase activity, which utilizes molecular oxygen and reducing equivalents from NAD(P)H;
- (d) oxidation, dealkylation, deaminoation, dehalogenation, and sulfoxide formation that are involved in a variety of biological events in plants and animals
10 (*e.g.,* catabolism, anabolism, and xenobiotic activities);
- (e) recognition of at two substrates: campestanol (CN) and 6-deoxocasterone (6-deoxoCS);
- (f) 22 α -hydroxylase activity;
- (g) DWF4 phenotypic activities such as modulation of cell length, periods of
15 flowering, branching, seed production, leaf size, and sterol composition in a plant;
- (h) regulation of gibberellic acid, cytokinins and/or auxin;
- (i) induce resistance to plant pathogens (*see, e.g.,* U.S. Patent No. 5,952,545);
- (j) accelerating growth at low temperatures; and
- (k) accelerating growth in dark conditions.

20 A DWF4 analog, whether a derivative, fragment or fusion of native DWF4 polypeptides, is capable of at least one DWF4 activity. Preferably, the analogs exhibit at least 60% of the activity of the native protein, more preferably at least 70% and even more preferably at least 80%, 85%, 90% or 95% of at least one activity of the native protein.

25 Further, such analogs exhibit some sequence identity to the native DWF4 polypeptide sequence. Preferably, the variants will exhibit at least 35%, more preferably at least 59%, even more preferably 75% or 80% sequence identity, even more preferably 85% sequence identity, even more preferably, at least 90% sequence identity; more preferably at least 95%, 96%, 97%, 98% or 99% sequence identity.

30 DWF4 analogs can include derivatives with increased or decreased activities as compared to the native DWF4 polypeptides. Such derivatives can include changes

within the domains, motifs and/or consensus regions of the native DWF4 polypeptide, which are described in detail in Example 3.

Once class of analogs is those polypeptide sequences that differ from the native DWF4 polypeptide by changes, insertions, deletions, or substitution; at
5 positions flanking the domain and/or conserved residues. For example, an analog can comprise (1) the domains of a DWF4 polypeptide and/or (2) residues conserved between the DWF4 polypeptide and other cytochrome P450 proteins, for example as shown in Figure 3 and described in Example 3.

Another class of analogs includes those that comprise a DWF4 polypeptide
10 sequence that differs from the native sequence in the domain of interest or conserved residues by a conservative substitution. For example, an analog that exhibits increased sterol binding can have optimized sterol binding domain sequences that differ from the native sequence.

Yet another class of analogs includes those that lack one of the *in vitro*
15 activities or structural features of the native DWF4 polypeptides, for example, dominant negative mutants or analogs that comprise a heme-binding domain but contain an inactivated steroid binding domain.

DWF4 polypeptide fragments can comprise sequences from the native or analog sequences, for example fragments comprising one or more of the following
20 P450 domains or regions: A, B, C, D, anchor binding, and proline rich. Such domains and regions are shown in Figures 2B, 3 and described in Example 3.

Fusion polypeptides comprising DWF4 polypeptides (*e.g.*, native, analogs, or fragments thereof) can also be constructed. Non-limiting examples of other polypeptides that can be used in fusion proteins include chimeras of DWF4
25 polypeptides and fragments thereof; and P450 polypeptides or fragments thereof, such as those shown in Figure 3.

In addition, DWF4 polypeptides, derivatives (including fragments and chimeric proteins), mutants and analogues can be chemically synthesized. See, *e.g.*, Clark-Lewis et al. (1991) *Biochem.* 30:3128-3135 and Merrifield (1963) *J. Amer.*
30 *Chem. Soc.* 85:2149-2156. For example, DWF4, derivatives, mutants and analogues can be synthesized by solid phase techniques, cleaved from the resin, and purified by preparative high performance liquid chromatography (*e.g.*, see Creighton, 1983,

Proteins, Structures and Molecular Principles, W. H. Freeman and Co., N.Y., pp. 50-60). DWF4, derivatives and analogues that are proteins can also be synthesized by use of a peptide synthesizer. The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (e.g., the Edman degradation
5 procedure; see Creighton, 1983, Proteins, Structures and Molecular Principles, W. H. Freeman and Co., N.Y., pp. 34-49).

Further, the *dwf4* polynucleotides and DWF4 polypeptides described herein can be used to generate antibodies that specifically recognize and bind to the protein products of the *dwf4* polynucleotides. (See, Harlow and Lane, eds. (1988)
10 "Antibodies: A Laboratory Manual"). The DWF4 polypeptides and antibodies thereto can also be used in standard diagnostic assays, for example, radioimmunoassays, ELISA (enzyme linked immunoradiometric assays), "sandwich" immunoassays, immunoradiometric assays, in situ immunoassay, western blot analysis, immunoprecipitationassays, immunofluorescent assays and PAGE-SDS.

15

Applications

The present invention finds use in various applications, for example, including but not limited to those listed above.

The polynucleotide sequences may additionally be used to isolate mutant *dwf4*
20 gene alleles. Such mutant alleles may be isolated from plant species either known or proposed to have a genotype which contributes to altered plant morphology. Additionally, such plant *dwf4* gene sequences can be used to detect plant *dwf4* gene regulatory (e.g., promoter or promoter/enhancer) defects which can affect plant growth.

25 The molecules of the present invention can be used to provide plants with increased seed and/fruit production, extended flowering periods and increased branching. The molecules described herein can be used to alter the sterol composition of a plant, thereby increasing or reducing cholesterol content in the plant. A still further utility of the molecules of the present invention is to provide a tool for
30 studying the biosynthesis of brassinosteroids, both *in vitro* and *in vivo*.

The *dwf4* gene of the invention also has utility as a transgene encoding a cytochrome P450 protein that mediates multiple 22 α hydroxylation steps in

brassinosteroid biosynthesis which results in a transgenic plant to alter plant structure or morphology. The *dwf4* gene also has utility for encoding the DWF4 protein in recombinant vectors which may be inserted into host cells to express the DWF4 protein. Further, the *dwf4* polynucleotides of the invention may be utilized (1) as
5 nucleic acid probes to screen nucleic acid libraries to identify other enzymatic genes or mutants; (2) as nucleic acid sequences to be mutated or modified to produce DWF4 protein variants or derivatives; (3) as nucleic acids encoding 22 α -hydroxylase in molecular biology techniques or industrial applications commonly known to those skilled in the art.

10 The *dwf4* nucleic acid molecules may be used to design plant *dwf4* antisense molecules, useful, for example, in plant *dwf4* gene regulation or as antisense primers in amplification reactions of plant *dwf4* gene nucleic acid sequences. With respect to plant *dwf4* gene regulation, such techniques can be used to regulate, for example, plant growth, development or gene expression. Further, such sequences may be used
15 as part of ribozyme and/or triple helix sequences, also useful for *dwf4* gene regulation.

The *dwf4* control element (*e.g.*, promoter) of the present invention may be utilized as a plant promoter to express any protein, polypeptide or peptide of interest in a transgenic plant. In particular, the *dwf4* promoter may be used to express a protein involved in brassinosteroid biosynthesis.

20 The Arabidopsis DWF4 protein of the invention can be used in any biochemical applications (experimental or industrial) where 22 α -hydroxylase activity is desired, for example, but not limited to, regulation of BL synthesis, regulation of other sterol synthesis, modification of elongating plant structures, and experimental or industrial biochemical applications known to those skilled in the art.

25

EXPERIMENTAL

Below are examples of specific embodiments for carrying out the present invention. The examples are offered for illustrative purposes only, and are not intended to limit the scope of the present invention in any way.

30 Efforts have been made to ensure accuracy with respect to numbers used (*e.g.*, amounts, temperatures, etc.), but some experimental error and deviation should, of course, be allowed for.

Example 1: Materials and Methods

A. Plant Growth Conditions

5 The conditions used for plant growth were essentially as described previously (Feldmann (1991) *Plant J.* 1:71-82 ; Forsthoefel et al. (1992) *Aust. J. Plant Physiol.* 19:353-366), except that agar-solidified medium contained 0.5% sucrose. Seedlings up to 2 weeks of age (6 weeks of age for dark-growth experiments) were grown on 0.8% agar-solidified medium containing 1 x Murashige and Skoog 1962 salts (Murashige, T., and Skoog, F. (1962) *Physiol. Plant.* 15:473-497) and 0.5% sucrose (w/v) and cold treated (4°C) for 2 days in the dark before transfer to the light (24 hr light; 80 $\mu\text{mol m}^{-2} \text{sec}^{-1}$); older plants were grown in potting soil. The plates were sealed with Parafilm (American National Can Co., Chicago, IL) for the entire experiment. For nucleic acid extraction, genetic analysis, and other experiments in which mature plants were required, seeds were sown on Metromix 350 (Grace Sierra, Milpitas, CA) presoaked with distilled water. The pots were covered with plastic wrap and cold treated (4°C) for two days before transfer to a growth chamber (16:8, light [240 $\mu\text{mol m}^{-2} \text{sec}^{-1}$]:dark; 22 and 21 °C, respectively, and 75 to 90% humidity). The plastic wrap was removed 5 days after germination, and the pots were subirrigated in distilled water as required. Germination of seeds for dark growth experiments was induced by overnight exposure of the seeds to light immediately after removing the plates from incubation at 4°C. The *dwf4-1* and *dwf4-2* mutations were in the *Arabidopsis thaliana* ecotype Wassilewskija (Ws-2) background; the *dwf4-3* and *dwf4-4* mutations were in the Enkheim (En-2) background.

25

B. Analytical Methods

Protoplasts were obtained by overnight incubation of sliced leaves in 0.1% cellulysin, 0.1% driselase, 0.1% macerasc (Calbiochem, San Diego, CA) in 125 mM Mes, pH 5.8, 0.5 M mannitol, and 7 mM CaCl₂ (Galbraith et al. (1992) *Planta* 186:324-326 . Immediately before observation, chloroplasts were stained with a solution of 1.5% KI and 1% I₂. Measurements were performed as described for tissue sections, and plane areas were calculated according to the formula $A = \pi r^2$.

30

Chlorophyll determinations were performed from 2-week-old soil-grown plants. Green tissue was weighed, frozen in liquid nitrogen, and extracted in dim light with 80% acetone in the presence of a mixture of equal parts sand, NaHCO_3 , and Na_2SO_4 . After brief centrifugation, the supernatant was collected and the extraction
5 was repeated twice, pooling the supernatants from each sample. Chlorophylls a and b were measured spectrophotometrically, as described in Chory et al. (1991), *supra*.

C. Growth Signal Response Measurements

Gibberellic acid (GA) response was assayed on plants grown individually in
10 5.7-cm pots. Once inflorescences reached 1 to 2 mm in height, they were sprayed weekly with 1 mM GA3 (Sigma). Control plants were sprayed with water. One week after the third spraying, plants were collected, and the length of the main stem was measured between the top of the rosette and the base of the most distal pedicel; 13 to 18 plants of each line were measured per treatment. Auxin response was tested by
15 growing seedlings for 10 days under 16 hr of light on vertically oriented agar plates containing various concentrations of 2,4-D (Gibco, Grand Island, NY). Genetic interaction with the *hy2* mutation was tested by growing seedlings under continuous light for 7 days. Brassinolide (BL) response was determined in liquid culture, as described by Clouse et al. (1993), *supra*, except that three or four seedlings were
20 grown in each well of a 24-well culture plate for 7 days. Measurements were taken for 10 to 20 seedlings for each genotype and condition, under a dissection microscope fitted with an ocular micrometer.

D. Microscopy

25 Tissues were fixed in 2% glutaraldehyde and 0.05 M sodium cacodylate, pH 6.9, for 2 hr at room temperature or overnight at 4°C, followed by three washes in buffer. For light microscopy, 1% safranin was included in the first wash, and embedding was performed in Paraplast Plus (Oxford Labware, St. Louis, MO). Ten-millimeter sections from five individual plants per line were analyzed and
30 photographed, and cell measurements were taken using a ruler on 5 x 7 inch prints. A print of a hemocytometer grid at the same final magnification was used for calibration. At least 25 cells were measured per sample, with a minimum of 150 cells

per line. For electron microscopy, the tissues were treated after fixation with 1% tannic acid in buffer for 30 min, washed three times, and postfixed in 1% OsO₄ in buffer for 2 hr, followed by five washes and dehydration through an ethanol series. Samples for transmission electron microscopy were embedded in Spurr's resin.

- 5 Sections (90 nm) were stained with saturated uranyl acetate followed by Reynolds's lead citrate (Reynolds (1963) *J. Cell Biol.* 17:208-212) and examined in a JEOL (Tokyo, Japan) 100-CX instrument. For scanning electron microscopy, samples were transferred to freon 113, critical point dried, and sputter-coated with 30 to 50 nm of gold. Analysis was performed in a microscope (ISI model DS130; Topcon, Inc.,
10 Paramus, NJ) with an accelerating voltage of 15 kV. Electron microscopy was performed at the Electron Microscope Facility, Division of Biotechnology, Arizona Research Laboratories, University of Arizona.

Example 2: Isolation of *dwf4* gene

15

A. Isolation of the DWF4 Gene

- The *dwf4-1* mutation was identified in a screen of 14,000 transformants of Arabidopsis, resulting in a dwarfed phenotype similar to *dwf1* (Feldmann and Marks (1987) *Mol. Gen. Genet.* 208:1-9; Feldmann et al. (1989) *Science* 243:1351-1354;
20 referred to as diminuto in Takahashi et al. (1995) *Genes Dev.* 9:97-107 and Szekeres et al., *supra*) and *det2* (Azpiroz et al. (1998), *supra*). Two independent lines were found that segregated for a similar phenotype: both were shorter than *dwf1*, but their rosette diameter was comparable to that mutant. These dwarfs were also essentially infertile. The most striking aspect of the morphology of these mutants is their
25 similarity to *det2* (Chory et al. (1991) *Plant Cell* 3:445-459). For this reason, further analysis was conducted with these lines. After being found to be allelic to each other, both were designated as *dwf4*.

- dwf4-1* segregated for a single kanamycin resistance marker, and gel blot analysis with DNA from single plants of this family confirmed that the pattern is
30 consistent with a single insert. The *dwf4* mutation was subsequently shown to be inherited as a monogenic, recessive Mendelian trait that, in *dwf4-1*, cosegregates with the dominant kanamycin resistance marker contained in the T-DNA, suggesting that

the mutation in this line may be a disrupted, tagged allele. *dwf4-2* also contains a single kanamycin resistance marker, but it failed to cosegregate with the dwarf phenotype. Two additional alleles (*dwf4-3* and *dwf4-4*) were identified among dwarf mutants obtained from the Nottingham Arabidopsis Resource Centre (Nottingham, UK; N365 and N374). Unless otherwise indicated, all experiments presented below were performed with *dwf4-1*.

Standard molecular techniques were performed as described previously (Sambrook et al. 1989). The plant DNA flanking the T-DNA was cloned using the plasmid rescue technique as described by Dilkes and Feldmann (1998) "Cloning genes from T-DNA tagged mutants" in *Methods in Molecular Biology: Arabidopsis Protocol*, J. Martinez-Zapater and J. Salinas, eds (Totowa, NJ: Humana Press), pp. 339-351. Briefly, *dwf4-1* genomic DNA was digested with EcoRI (for the right border) or SalI (for the left border), ligated under conditions to maximize intramolecular events, and introduced into competent *Escherichia coli* cells. The resulting colonies were screened on ampicillin. Five colonies from the left border transformation contained plant DNA flanking the insertion site. The restriction pattern displayed two different types of plant DNA. Three contained a 5.6-kb insert, whereas the other two contained a 1.1-kb insert. This result suggested that the T-DNA insert in *dwf4-1* was flanked by two left border sequences. The existence of two left border sequences was confirmed by gel blot analysis with genomic DNA, using the putative plant flanking DNAs as probes. A single wild-type EcoRI fragment was split into two fragments in *dwf4-1*.

Wild-type genomic clones were isolated from a library made from Ws-2 DNA by using the 5.6-kb fragment as a probe. The library was constructed using λ DASH-II arms (Stratagene, La Jolla, CA). Approximately 10,000 primary plaques were screened. Duplicate-filter screening resulted in 12 positives. Restriction mapping of the secondary clones revealed that some contained part of the *DWF4* locus. In fact, one of the clones, D4G12-1, contained an intact 13-kb DNA spanning the T-DNA insertion site. The 13-kb insert in D4G12-1 was subcloned into pBluescript SK- (Stratagene). Subclones were sequenced from each end of the insert by using the universal primers in the plasmid. DNA sequencing was performed using an ABI 377 (Perkin-Elmer, Norwalk, CT) automated sequencer at the Arizona

Research Laboratories (Tucson, AZ).

Reverse transcriptase-polymerase chain reaction (RT-PCR) was used to isolate a cDNA clone. RNA was isolated from 5-day-old dark- and light-grown seedlings. Superscript II reverse transcriptase (BRL, Gaithersburg, MD) was used for the cDNA
5 synthesis, according to the manufacturer's protocol. Briefly, 7 µg of total RNA was mixed with the reverse primer, D4R3. To the heat-denatured RNA-primer mix, the RT mixture was added and incubated for 1 hr at 43°C. Two microliters of RT product was used for PCR amplification by using different primers sets intended to cover all of the putative coding region. RT-PCR products were fractionated on an 0.8%
10 agarose gel (Sambrook et al. 1989); the expected bands were purified using a GeneClean kit (BIO 101, Inc., Vista, CA), further amplified, and sequenced to determine the coding region.

B. Sequencing

15 *dwf4-2* was isolated from a T-DNA mutant population as an untagged allele, whereas *dwf4-3* and *dwf4-4* were obtained from plants obtained from the Nottingham Arabidopsis Stock Centre (University of Nottingham, UK; stock nos. N365 and N374); the mutagenesis method for these two lines is not known. Based on the DNA sequence of wild-type genomic DNA, pairs of primers were designed to amplify
20 ~1-kb stretches of genomic DNA. Oligonucleotide sequences are shown 5' to 3'. The numbers shown correspond to positions in the genomic sequence, with the adenine base in the translation initiation codon set as position 1. D4OVERF,
1-ATGTTTCGAAACAGAGCATCATACT-24 (SEQ ID NO:3); D4PRM,
(-1)-CCTCGATCAAAGAGAGAGAGA-(-21) (SEQ ID NO:4); D4RTF,
25 143-TTCTTGGTGAAACCATCGGTTATCTTAAA-171 (SEQ ID NO:5); D4RTR,
853-TATGATAAGCAGTTCCTGGTAGATTT-828 (SEQ ID NO:6); D4F1,
(-242)-CGAGGCAAC-AAAAGTAATGAA-(-222) (SEQ ID NO:7); D4R1,
689-GTTAGAAACTCTAAAGATTCA-669 (SEQ ID NO:8); D4F2,
576-GATTCTTGGCAACAAAACCTCTAT-598 (SEQ ID NO:9); D4R2,
30 1685-CCGAACATCTTTGAGTGCTT-1666 (SEQ ID NO:10); D4F3,
1606-GTGTGAAGGTTATAAATGAACTCTT-1631 (SEQ ID NO:11); D4R3,

3156-GGTTTAATAGTGTGCGACACTAATA-3132 (SEQ ID NO:12); D4F4,
2316-CCGATGACTTGTACGTGCGTTA-2337 (SEQ ID NO:13); D4F5,
730-GCGAAGCATATAATGAGTATGGAT-753 (SEQ ID NO:14); and D4R5,
1876-GTTGGTCATAACGAGAATTATCCAAA-1851 (SEQ ID NO:15). Because

5 the two stock center lines were in a different genetic background than the wild-type
gene that we had sequenced (WS), primers were based primarily on the exon sequence
to avoid sequence variation between introns. Genomic DNA isolated from the
mutants was subjected to PCR, using these primer sets. The amplified DNA
fragments were fractionated on 0.8% TAE agarose gel (Sambrook et al. 1989),
10 purified using GeneClean (BIO 101, Inc.) or Qiaquick™ columns (Qiagen Inc.,
Chatsworth, CA), and sequenced. Putative mutations were identified by comparing
the mutant DNA sequence with the wild-type sequence. The sequence was confirmed
by sequencing independently amplified fragments at least three times for each
mutation to eliminate PCR misincorporation.

15

C. Sequence Analysis

Annotations in multiple sequence alignment were performed using the
ALSCRIPT package provided by Barton, G.J. (1993) *Protein Eng.* 6:37-40. Searches
for similar protein sequence were performed with the BLAST program (Altschul et al.
20 (1990), *supra*). In addition, useful packages, available on the internet, such as
promoter, protein targeting, polyadenylation site, and splice site, have been employed
to characterize the DNA and protein sequence (consolidated in the search launcher,
Baylor College of Medicine, Baylor, TX). All other sequence analysis was performed
using the Genetics Computer Group (Madison, WI) software package.

25 Analysis of the complete genomic sequence, starting at the EcoRI site, with
the promoter prediction by neural network (NNPP) package
(<http://www-hgc.lbl.gov/projects/promoter.html>), indicated that the gene included a
putative promoter (TATAT is found in the putative promoter region between
nucleotides -143 to -78) and polyadenylation signal sequences (AATAA near a
30 position at 3238 bp and a putative GU-rich signature from 3283 to 3290 bp).

Unsuccessful attempts to detect mRNA by tissue-specific RNA gel blot
analysis, using the 4.8-kb fragment as a probe, suggested that *DWF4* encoded a rare

message. In addition, there were no matching expressed sequence tags in the Arabidopsis database. Therefore, we screened two different cDNA libraries made with either normalized mRNA from different tissues or RNA from floral tissues, using the 4.8-kb fragment as a probe (ABRC stock numbers CD4-7 and CD4-6, 5 respectively). After finding no positives in 109 clones screened, we chose to directly amplify DWF4 cDNA from total RNA made from 5-day-old seedlings, using reverse transcriptase-polymerase chain reaction (RT-PCR). Whereas RNA from both light-grown and dark-grown seedlings yielded the expected RT-PCR products, RNA from dark-grown seedlings generated significantly more. The bands were gel purified 10 and sequenced. Alignment of the genomic and cDNA sequences indicated that the *DWF4* gene was composed of eight exons and seven introns (Figure 2A; Figure 10).

Sequence analysis of the *dwf4-1* allele revealed that the T-DNA was inserted in the 5' end of intron 7 (Figure 2A). In addition, sequence analysis of the left border plant junctions indicated that at one junction (5'), 75 bp of unknown DNA was 15 inserted, whereas at the other junction (3'), 24 bp of left border and 19 bp of plant DNA were deleted. To prove that DWF4 had been cloned, two other *dwf4* alleles (*dwf4-2* and *dwf4-3*) were sequenced to identify possible lesions. As shown in Figure 2B, *dwf4-2* contained a deletion of three conserved amino acids (324 to 326) caused by a 9-bp deletion, and *dwf4-3* contained a premature stop codon (289) caused by 20 changing a tryptophan codon (UGG) to a nonsense codon (UGA). Due to a premature stop codon, translation is predicted to be terminated before the heme binding domain, which is essential for cytochrome P450 function (Poulos et al. (1985) *J. Biol. Chem.* 260:16122-16130). Because T-DNA-generated alleles *dwf4-1* and *dwf4-2* and an additional mutant allele all possess loss-of-function mutations affecting the same 25 protein, we conclude that we have cloned the *DWF4* gene.

Example 3: The *DWF4* Gene Encodes a Cytochrome P450

The open reading frame of *DWF4* encodes a protein composed of 513 amino acids. BLAST database searches (Altschul et al. (1990) *J. Mol. Biol.* 215:403-410) 30 for similar sequences yielded a superfamily of cytochrome P450 proteins as significant high-scoring segment pairs. Cytochrome P450s are heme-thiolate enzymes. They display a characteristic Soret absorption peak at 450 nm when the

substrate-bound, reduced form is exposed to the light (Jefcoate (1978) "Measurement of substrate and inhibitor binding to microsomal cytochrome P-450 by optical-difference spectroscopy" in *Methods in Enzymology*, Vol. 52, S. Fleischer and L. Packer, eds (London: Academic Press), pp. 258-279). Typical microsomal

5 cytochrome P450s hydroxylate various substrates via their monooxygenase activity, which utilizes molecular oxygen and reducing equivalents from NAD(P)H. In addition to the hydroxylation, other activities of cytochrome P450 enzymes, such as oxidation, dealkylation, deamination, dehalogenation, and sulfoxide formation, are involved in a variety of biological events in catabolism, anabolism, and xenobiotic

10 metabolism in plants as well as animals (reviewed in West (1980) "Hydroxylases, monooxygenases, and cytochrome P-450" in *The Biochemistry of Plants: A Comprehensive Treatise*, Vol. 2, Metabolism and Respiration, D.D. Davies, ed (New York: Academic Press), pp. 317-365; Nebert and Gonzalez (1987), *supra*; Guengerich (1990) *Crit. Rev. Biochem. Mol. Biol.* 25:97-152, Guengerich (1993) *Am. Sci.*

15 81:440-447; Durst (1991) "Biochemistry and physiology of plant cytochrome P-450" in *Microbial and Plant Cytochromes P-450: Biochemical Characteristics, Genetic Engineering and Practical Implications*, K. Ruckpaul and H. Rein, eds (London: Taylor and Francis), pp. 191-232; Bolwell et al. (1994) *Phytochemistry* 37:1491-1506; Durst and Nelson (1995), *supra*; Schuler (1996) *CRC Crit. Rev. Plant*

20 *Sci.* 15:235-284). Evolutionarily, cytochrome P450s have been found in a broad spectrum of living organisms, and they share significant homology at the amino acid sequence level. Thus, it has been proposed that all known cytochrome P450s were derived from a common ancestor (Nelson and Strobel (1987) *Mol. Biol. Evol.* 4:572-593).

25 Typical cytochrome P450s contain four characteristic domains as defined by Kalb and Loper 1988. Of the four domains, A, B, C, and D, at least two of them have been assigned specific functions. Domain A binds a substrate and molecular oxygen, and domain D has been shown to bind heme-prosthetic groups via a thiolate bond (Poulos et al. 1985). Thus, typically, microsomal cytochrome P450 enzymes can be

30 identified by their characteristic signature sequences, including the heme binding domain, domain A (also referred to as dioxygen binding), domain B (steroid binding), and domain C (Nebert and Gonzalez (1987) *Annu. Rev. Biochem.* 56:945-993; Kalb

and Loper (1988) *Proc. Natl. Acad. Sci. USA* 85:7221-7225). All of these signature sequences were found in DWF4; the relative positions of the domains are indicated in Figure 2B.

Durst and Nelson (1995) *Drug Metab. Drug Interact.* 12:189-206 classified
5 plant cytochrome P450s into two distinct groups based on their clustering nature in a phylogenetic tree. All of the group A families cluster and are assumed to originate from a common plant P450 ancestor. The group A cytochrome P450s conform to the characteristic consensus sequences (A/G)GX(D/E)T(T/S) in domain A (also called helix I) and PFG(A/S/V)GRRXC(P/A/V)G of the heme binding domain (D) with only
10 a few exceptions. Group A cytochrome P450s appear to catalyze plant-specific reactions such as lignin biosynthesis (Figure 6; GenBank accession number P48421). By contrast, P450s that do not belong to group A (non-A P450s) are scattered in the phylogenetic tree. They share more amino acid identity/similarity with P450s found in animals, microbes, and fungi than with those found in plants. The non-A P450s
15 possess functions, such as steroid metabolism, that are not limited to plants. Generally, non-A P450s have limited homology with known domains described for group A.

The most similar protein to DWF4 is the Arabidopsis CPD protein, a non-A P450. A mutation in *CPD* also caused dwarfism (Szekeres et al. 1996; CYP90A1,
20 GenBank accession number X87368). DWF4 and CPD share 43% identity and 66% similarity. Conforming to the recommended nomenclature for cytochrome P450 enzymes, DWF4 and CPD (CYP90A1) are grouped into the same family within different subgroups (Durst and Nelson (1995) *Drug Metab. Drug Interact.* 12:189-206). As such, DWF4 represents a second member of the CYP90 family and
25 is designated CYP90B1. Sequence similarity between the two proteins occurs throughout their length, with the greatest similarity in the classically conserved domains. Residues conserved between DWF4 and CYP90A are boxed and italicized in Figure 3. The second most similar protein is the tomato CYP85 (Bishop et al. (1996), *supra*; GenBank accession number U54770). A mutation in this gene also
30 results in dwarfism. DWF4 and CYP85 share 35% identity and 59% similarity in their overall protein sequences.

Six cytochrome P450 sequences with the greatest homology to DWF4, CYP90A1, CYP85, CYP88 (Winkler and Helentjaris (1995) *Plant Cell* 7:1307-1317; GenBank accession number U32579), cyanobacteria CYP120 (Kaneko et al. (1996) *DNA Res.* 3:109-136; GenBank accession number D64003), human CYP3A3X (Molowa et al. (1986) *Proc. Natl. Acad. Sci. USA* 83:5311-5315; GenBank accession number M13785), and zebrafish CYP26 (White et al. White (1996) *J. Biol. Chem.* 271:29922-29927; GenBank accession number U68234), were chosen for multiple sequence alignment. Putative domains defined by Kalb and Loper (1988), *supra* are boxed and labeled in Figure 3. First, the heme binding domain pFGgFpRlCpGkel matches completely the sequence defined previously. Uppercase letters in the domain indicate amino acids conserved at all seven sequences in the alignment, and lower-case letters represent residues conserved in at least half of the proteins. Of the amino acids conserved in the heme binding domain, the function of the cysteinyl is established as a thiolate ligand to the heme (Poulos et al. (1985), *supra*).

Domain A is defined by xllfaGhEttssxIxxa. Lowercase x's indicate variable amino acids. An invariant glutamate (E) preceded threonine (T) at position 314, T314, which is believed to bind dioxygen, was conserved in all proteins compared except CYP88 of maize. The second signature sequence, domain B, is also conserved in *DWF4* with significant similarity. A valine at position 370 is conserved in all of the proteins, but it does not appear in Kalb and Loper's classic report (1988) on conserved domains. Again, *DWF4* matches the domain C consensus sequence. Finally, the anchoring domain in the N-terminal end was distinguished by a repeat of the hydrophobic residue leucine. In addition, in *DWF4*, two acidic (glutamate) and two basic (histidine) residues precede the repeated leucine in the N-terminal leader sequence. These charged residues may add more stability to the membrane topology of the protein as a strong start-stop transfer peptide (von Heijne (1988) *Biochim. Biophys. Acta* 947:307-333).

Thus, phylogenetic analyses of these seven proteins with cytochrome P450s unique to plants (group A; Durst and Nelson (1995), *supra*) indicate that *DWF4* does not cluster with these cytochrome P450s (Figure 6). Rather, *DWF4* clustered with cytochrome P450s from other organisms: cyanobacteria (CYP120), rat (CYP3A2), human (CYP3A3X), and plants (CYP90, CYP85, and CYP88). *DWF4* also deviates

from the consensus sequence in the group A heme binding domain in that it possesses a PFGGGPRLCAG sequence in which arginine (R) is substituted for proline (P). However, domain A of DWF4, AGHETS, fits the consensus of domain A of group A. These characteristics suggest that DWF4 is a monooxygenase, similar to P450s of group A, that utilizes molecular oxygen as a source of the hydroxyl group, but it
5 mediates some reaction(s) that are not necessarily specific for plants, for instance, steroid hormone biosynthesis, which is a critical event for animals. In fact, the similarity of DWF4 to the rat testosterone 6 β -hydroxylase (34%; GenBank accession number 631895) or glucocorticoid-inducible hydroxylase (31%; Molowa et al. 1986;
10 GenBank accession number M13785) supports this idea. Further, the similarity that DWF4 shares with CYP90A and CYP85, 66 and 59%, respectively, is additional proof that it is involved in plant steroid biosynthesis (Bishop et al. 1996 ; Szekeres et al. 1996).

15 **Example 4: The *dwf4* Phenotype**

As formally defined, a plant with a dwarf phenotype is one that has a short, robust stem and short, dark green leaves. *dwf4* mutants are significantly smaller than the wild type and are dark green in color. They have short, rounded leaves. Again, the *dwf4* phenotype is reminiscent of the light-regulatory mutant *det2* (Chory et al.,
20 *supra*); however, complementation analysis has shown that the two mutations are not allelic, with the *dwf4* mutation mapping to the lower arm of chromosome 3 and *det2* mapping to chromosome 2 (Chory et al., *supra*). The results presented in Table 1 show that soil-grown *dwf4* plants attained a height of <3 cm at 5 weeks, whereas wild-type plants grew to >25 cm. Moreover, individual organs, such as leaves, were
25 invariably shorter in dwarf plants. *dwf4* siliques were also markedly shorter than those of the wild type and were infertile. The loss of fertility of *dwf4* was due to the reduced length of the stamen filaments relative to the gynoecium, which resulted in mature pollen deposition on the ovary wall rather than on the stigmatic surface. Hand pollination of *dwf4* flowers with either mutant or wild-type pollen resulted in good
30 seed set without significantly changing the size of the siliques.

Table 1: The Development of Wild-Type and <i>dwf4-1</i> plants		
Measurement	Wild-Type ^a	<i>dwf4-1</i> ^a
Five Weeks		
Height	25.8 ± 2.6 cm	2.8 ± 0.3 cm
Leaf blade length ^b	1.72 ± 0.36 cm	0.96 ± 0.15 cm
Leaf blade width ^b	0.77 ± 0.10 cm	0.99 ± 0.18
No. inflorescences	3.6 ± 0.5	10.5 ± 1.4
No. rosettes	7.1 ± 0.9	13.5 ± 1.3
Other		
start of flowering	21.5 days	25.9 days
mature silique length	1.16 ± 0.07 cm	0.29 ± 0 cm
No. seeds per silique	37.7 ± 3.3	0.0
Final no. of siliques	336.5 ± 90.6	988.4 ± 214.2
Height at maturity	27.0 ± 2.7	11.6 ± 1.0 cm

^a results shown are the average ±SD of measurements taken from 10 plants

^b measurements taken from the second pair of leaves

Another feature of *dwf4* plants is a reduction in apical dominance, as was evident by the threefold increase in the number of inflorescences at 5 weeks of age (Table 1). Mutants also had twice the number of rosette leaves, which may be explained by a prolonged vegetative phase in the *dwf4* plants. Development of flowers on the primary inflorescence was delayed by ~4 days in *dwf4*, but the flowering phase was significantly longer in the mutant, with senescence of the last flower occurring at ~98 days compared with ~57 days for the wild type. One result of this delay in senescence was that *dwf4* plants contained almost three times the number of siliques as did the wild type (Table 1).

The reduced stature observed in soil-grown *dwf4* was also observed in hypocotyls of agar-grown seedlings. Measurements of hypocotyl length over time

indicated that not only were *dwf4* seedlings shorter than wild-type seedlings immediately after germination but also that the rate of growth was retarded in the mutants (Figure 5). In addition, *dwf4* hypocotyls reached their terminal length in <5 days, whereas wild-type seedlings continued to grow.

5 In sum, the *dwf4* phenotype can be described as being due to both primary and secondary effects of reduced cell elongation. The primary effect is simply a reduction in the length of individual organs exclusively along their normal growth axis; that is, organ width is not reduced (Table 1). The secondary effects of reduced cell elongation are themselves due to the reduction in organ length. The dark green color of the
10 leaves, for example, may be due exclusively to the existence of a wild-type number of chloroplasts in a significantly smaller cell. Similarly, the sterility of mutants is a consequence of the shortness of the stamens, which fail to deposit their pollen on the stigmatic surface. In addition to the morphological alterations of *dwf4*, mutants display delayed development, the first sign of which occurs at flowering (Table 1).
15 Because rosette leaves are produced continuously during vegetative development, delayed flowering results in *dwf4* rosettes having almost twice the number of leaves observed in the wild type.

Example 5: The Growth Defect of *dwf4* Is Due to a Reduction in Cell Length

20 Both the short stature and the reduced growth rate of *dwf4* could be due to a defect in cell division or cell elongation or both. To distinguish between these possibilities, we analyzed sections from 7-day-old hypocotyls and 5-week-old inflorescence stems, by light microscopy, as described in Example 1. To minimize variations due to the developmental stage of the sample, we always took the stem
25 sections from the fourth internode. As shown in Table 2, the average cell size in *dwf4* is significantly smaller than in wild-type plants, whereas no differences were detected in the number of cells along the length of either organ between the wild type and *dwf4*. Therefore, the short stature and reduced organ length of *dwf4* are largely or
30 observed in the number of cell layers contained in the wild type and *dwf4*.

Table 2: Cell Length in Wild-Type and <i>dwf4</i> plants		
Measurement	Wild-type	<i>dwf4</i>
average cell length in hypocotyl: 7 day old plant	92.7 μm	32.2 μm
average cell length in stem: 5-week old plant	79.2 μm	15.0 μm

The small size of *dwf4* cells offers a possible explanation for the dark green color of the mutant plants. Chlorophyll measurements were taken, and leaf mesophyll protoplasts were prepared, stained, and measured to visualize and count chloroplasts, as described in Methods. Although there were no significant differences in total chlorophyll content, the chlorophyll *a/b* ratio, or the absorption spectra between wild-type plants and mutants, the mean plane area (the apparent two-dimensional surface area of mounted cells) of *dwf4* leaf mesophyll protoplasts was 376 mm², whereas that of wild-type protoplasts was 599 mm². The two-dimensional comparison of plane area represents a dramatic reduction in volume for *dwf4* cells. However, the number of chloroplasts per cell was only slightly lower: the mean number of chloroplasts per cell was 40 for *dwf4* and 44 for the wild type. Therefore, *dwf4* cells contain a greatly increased number of chloroplasts per unit cell volume. As a consequence, the chloroplasts are brought closer to each other, making the color of the leaves appear darker. Chloroplast size was the same in both lines.

Thus, the rate of growth was significantly reduced in agar-grown *dwf4* seedlings, which ceased to grow when their hypocotyl length was <20% of the final wild-type length. Because all of the cells in a hypocotyl before the initiation of leaf development are present in the embryo, the initial growth of seedlings is due exclusively to cell expansion, which therefore must be reduced in *dwf4*. A similar situation applies to soil-grown plants. Five weeks after germination, well after plants had bolted, *dwf4* plants were shorter than wild-type plants (Table 1). Although the mutants continued growing for several weeks more than did the wild type, they remained shorter through senescence. That cell elongation is the direct cause of this decreased growth is shown by measurements of cell length both in 7-day-old hypocotyls (Table 2) and in 5-week-old stems (Table 2). Not only is the reduction in

cell length in good agreement with the reduction in organ length, but insofar as could be determined, there is no difference in the number of cells between *dwf4* and wild-type plants.

Organ growth by cell elongation in plants occurs as part of normal
5 development in response to a variety of input signals. Mutants that are defective in these signaling pathways invariably fail to elongate normally in response to the appropriate stimuli. A mutant with a block at a step that is common to several individual pathways would therefore be expected to have defective responses to all of the corresponding signals. *dwf4* appears to be such a mutant. Figure 6 shows that
10 elongation induced by the *hy2* mutation is blocked in a *dwf4 hy2* double mutant. Not surprisingly, in view of this result, *dwf4* also failed to display hypocotyl elongation as a response to growth in complete darkness. In addition, *dwf4* was capable of perceiving GA, but its response was severely compromised. This mutant could also respond to the inhibitory effects of auxin but was incapable of auxin-stimulated
15 elongation. It was only exogenous BL that fully restored wild-type length to *dwf4* hypocotyls (Choe et al. (1998), *supra*).

Because *dwf4* failed to respond to at least three independent signaling pathways but responded fully to only one, the most likely explanation for the dwarf phenotype is therefore that a fully functional BR system is required for a full response
20 to GA, auxin, and deetiolation. From the perspective of cellular economy, it may be advantageous that the downstream elements involved in cell elongation are shared among at least some of the signaling pathways that evoke this response. The interaction of various pathways at a common step provides the plant with a potential point for the integration of signals produced by diverse independent stimuli. Our
25 results indicate that BRs act at this downstream step.

Example 6: *dwf4* Is Specifically Rescued by Brs

The reduced length of cells in *dwf4* hypocotyls and inflorescence stems is indicative of a failure of these cells to elongate during development. A variety of
30 endogenous and environmental signals is responsible for stimulating elongation in plants; therefore, a series of experiments was performed to determine whether *dwf4* is

affected in a specific signaling pathway or is blocked in elongation as a response to various signals.

Of the endogenous (hormonal) signals that might be deficient in dwarf plants, an obvious candidate is GA, because gibberellin-deficient mutants are shorter in stature than are the wild-type plants (Koornneef and Van der Veen, *supra*). Our results, however, indicate that *dwf4* is not defective in the synthesis of gibberellins. When germinated on 10^{-5} M GA, wild-type seedlings demonstrated an elongation response (Figure 6), whereas *dwf4* seedlings responded minimally, if at all. At 10^{-4} M GA, wild-type seedlings elongated slightly more than at 10^{-5} M, but the *dwf4* seedlings were essentially saturated for elongation at 10^{-5} M GA. Similar results were obtained when soil-grown plants were sprayed with 1 mM GA once inflorescences first became visible: *dwf4* inflorescence stems elongated by only 28% above the untreated controls, whereas those of the wild type elongated by 45% above the untreated controls. Mutants that owe their reduced stature to decreased levels of endogenous gibberellins can be fully rescued by added hormone (Koornneef and Van der Veen, *supra*; Talon et al., *supra*). In addition, *dwf4* seeds germinate in the absence of exogenously supplied GA. Our results therefore suggest that *dwf4* is not deficient in endogenous GA. A corollary conclusion from this experiment is the demonstration that *dwf4* is capable of detecting GA; that is, it is not likely to be affected in signal perception but rather is defective in the extent to which it can respond to this signal.

Auxin can also stimulate cell elongation. This effect is especially visible in young seedlings (Klee and Estelle, *supra*). The response of wild-type and *dwf4* plants to auxin was tested by growing seedlings for 10 days on vertically oriented plates containing various concentrations of the synthetic auxin 2,4-D. At all concentrations assayed, inhibition of root growth was evident. Figure 6 shows that at 10^{-8} M 2,4-D, hypocotyl elongation in wild-type and *dwf4* seedlings was similar to that of the controls. Higher concentrations of auxin were inhibitory for both wild-type and *dwf4* seedlings, and lower concentrations had no effect. In view of the inhibition of root growth, it is clear that *dwf4* is not auxin resistant; rather, its elongation response is compromised.

As mentioned above, the most obvious exogenous signal for plants is light. Therefore, to investigate whether light-regulated cell elongation is altered in *dwf4*,

wild-type and *dwf4* seedlings were grown in the dark, as described in Example 1.

Figure 6 shows that as expected, wild-type seedlings displayed hypocotyl elongation typical of etiolated growth. By contrast, dark-grown *dwf4* seedlings were only slightly longer than those grown in the light. To assess the relationship between the

5 *dwf4* phenotype and light sensing by *dwf4*, the mutation was crossed into a mutant defective in the *HY2* gene. All *hy* mutants share the common phenotype of an elongated hypocotyl that mimics part of the etiolation response in the light. Specifically, *hy2* is deficient in active phytochrome because chromophore biosynthesis does not take place (Chory et al. (1989a) *Plant Cell* 1:867-880). Figure 6
10 shows that *dwf4 hy2* double mutants displayed a dwarfed phenotype indistinguishable from that of *dwf4 HY2* (light-grown control); therefore, the elongation block due to the *dwf4* mutation is epistatic to a defect in phytochrome activity.

In the course of our studies, we prepared a genomic library from *dwf4-1*, from which we isolated a clone in which a fragment of T-DNA interrupts a gene encoding a
15 putative cytochrome P450 steroid hydroxylase. Because BRs have been shown to elicit elongation in Arabidopsis (Clouse et al. (1993) *J. Plant Growth Regul.* 12:61-66) and because BR-deficient mutants have been recently described (Kauschmann et al. (1996), *supra*; Li et al. (1996), *supra*; Szekeres et al., *supra*), we tested the effect of BL on Arabidopsis seedlings by germinating seeds in liquid
20 medium containing different amounts of BL. As shown in Figure 6, the *dwf4* hypocotyls were restored to wild-type height by 10^{-6} M BL. This, together with our identification of a disrupted gene encoding a putative BR biosynthetic enzyme, strongly suggests that the phenotype of *dwf4* is specifically due to a defect in BR biosynthesis (see Choe et al. (1998) *Plant Cell* 10:231-243).

25 Thus, the results indicate that BL is involved at or near a downstream control point where multiple signaling pathways interact. First, as shown in Figure 6, BL is required for cell elongation as a response to darkness as well as GA and auxin. In addition, previous studies (Kauschmann et al. (1996), *supra*; Li et al. (1996), *supra*; Szekeres et al. (1996), *supra*) and the work described herein show that BR can
30 compensate for the cell elongation defect of mutants as diverse as *det2*, *cpd*, *dwf4*, *det1*, *cop1*, and *dwf1*. This places BRs downstream of all the cellular functions affected in these mutants. Finally, at least one of the BR biosynthetic genes has been

shown to be modulated by light, cytokinins, and the carbon source (Szekeres et al. (1996), *supra*).

Mutations in *axr2* result in a dwarf growth habit and a dark-grown phenotype with short hypocotyl and open cotyledons (Timppe et al. (1992), *supra*). In addition, *axr2* mutants are resistant to auxin, ethylene, and abscisic acid and have defective root and shoot gravitropism. The dwarf phenotype in *axr2* mutants has been shown to be due to reduced cell elongation and is rescued by BL (Szekeres et al. (1996), *supra*). This suggests that at least one of the multiple hormone signaling pathways affected in *axr2* involves a BR-dependent step. Mutations at another locus, *acaulis1*, also have a significant reduction in cell elongation, but the defect is confined to inflorescence stems and leaves (Tsukaya et al. (1993) *Development* 118:751-764). Flowers are fully fertile and mature into normal-sized siliques with normal seed set. There is no change in hypocotyl length. If BRs are directly involved in this apparently organ-specific signaling pathway, it may be due to organ-specific responsiveness to individual BR species. With regard to the mechanism of action of BRs, at the moment one can only speculate that the target may be a component of the cell expansion machinery. Perhaps steroid signaling initiates a series of events leading to cell wall loosening.

Example 7: The Elongation Defect of *dwf4* Leads to a Light-Regulatory Phenotype

The BR-deficient mutant *det2* was originally identified as defective in regulation by light (Chory et al. (1991), *supra*). Given the similarity of *det2* and *dwf4* phenotypes and functions and in view of the observation that *dwf4* is epistatic to *hy2*, one can predict that the etiolation response, which includes significant hypocotyl elongation, would not be normal in *dwf4*. To assess to what extent the etiolation response is affected by BR-dependent cell elongation, we grew *dwf4* and wild-type plants on agar under continuous light or in complete darkness, as described above in Example 1. After 7 days of growth in the light, wild-type seedlings displayed open and expanded cotyledons as well as emerging leaf buds. In contrast, the overall appearance of light-grown *dwf4* seedlings was strikingly similar to that of *det2* (Chory et al. (1991), *supra*). *dwf4* hypocotyls were very short, and the cotyledons were smaller than those of the wild type, displaying significant epinastic growth. As

expected, dark-grown wild-type seedlings had a typical etiolated appearance, with a highly elongated hypocotyl and closed, unexpanded cotyledons. However, *dwf4* hypocotyls failed to elongate. That the *dwf4* mutation can abolish the elongation component of the etiolation response is in agreement with the notion that the block in
5 cell elongation in *dwf4* is specifically a BR-dependent process.

In addition to short hypocotyls, dark-grown *dwf4* seedlings displayed partially open cotyledons and leaf primordia, with up to four leaf buds clearly visible. This has not been observed with the wild type, although it occurs with certain light-regulatory mutants (Chory et al. (1989b), *supra*; Deng et al. (1991) *Genes Dev.* 5:1172-1182;
10 Wei and Deng (1992), *supra*). *dwf4* leaf development continued in the darkness for several weeks, resulting in significant expansion of rosette leaves. These results indicate that *dwf4* plants can initiate what is normally a photomorphogenic pathway in the absence of light. Although this is often diagnostic of a light-regulatory mutant, wild-type *Arabidopsis* can perform leaf development and even flowering in complete
15 darkness when grown in liquid culture (Araki and Komeda (1993) *Plant J.* 4:801-811).

The cause for this dark-flowering effect is not understood; therefore, the possibility exists that leaf development in dark-grown *dwf4* is related to dark flowering and not to a light-regulatory defect. For example, perhaps the proximity of
20 the *dwf4* shoot apical meristem to the surface of the agar, due to the shortness of the hypocotyls, mimics some effect of submerged culture, such as a high water potential or a high concentration of some nutrient. To test this possibility, wild-type seedlings were grown in complete darkness for 6 weeks in vertically oriented dishes to maximize contact between the seedling and the medium. Wild-type seedlings grown
25 in this fashion displayed open cotyledons and underwent at least partial leaf development. In fact, all wild-type seedlings grown along the surface of the agar showed development of an inflorescence with at least one cauline leaf and a terminal flower bud. We conclude, therefore, that the appearance of leaves in dark-grown *dwf4* may be due simply to its short size and the culture conditions.

30 A number of light-regulatory mutants have been described that undergo photomorphogenesis in the dark at the cellular level. In mutants such as *cop1*, *cop8*, *cop9*, *cop10*, and *cop11*, stomata undergo photomorphogenic maturation (Deng and

Quail (1992), *supra* ; Wei and Deng (1992), *supra* ; Wei et al. (1994), *supra*); of these, *cop1* and *cop9* as well as *det1* (Chory et al. (1989b), *supra*) also initiate differentiation of plastids into chloroplasts. To determine whether *dwf4* plants undergo photomorphogenic cellular differentiation in the dark, we analyzed

5 cotyledons from light- and dark-grown plants by transmission and scanning electron microscopy. Analysis of plastids in thin sections from 7-day-old dark-grown seedlings showed no difference between the wild type and *dwf4*. Both lines contained normal chloroplasts when grown in the light, whereas dark-grown seedlings contained etioplasts, with their characteristic prolamellar body and no significant organization of

10 thylakoids. Analysis of stomatal structures on the underside of cotyledons from 7-day-old seedlings indicates that stomatal development was not completed in the dark, because the stomatal opening was occluded in both lines. The majority of light-regulatory mutants analyzed to date displayed light-grown morphology in the dark without concomitant chloroplast or stomatal development. As in these mutants,

15 therefore, the *dwf4* mutation uncouples the developmental pathway of seedling morphology from that of light-regulated cellular differentiation.

An additional feature of many light-regulatory mutants is that photomorphogenesis in the dark is accompanied by expression of genes that normally are light induced (Chory et al. (1989b), *supra* , Chory et al. (1991), *supra* ; Deng et al.

20 (1991), *supra* ; Wei and Deng (1992), *supra* ; Hou et al. (1993), *supra* ; Wei et al. (1994), *supra*). To assess whether *dwf4* is able to induce light-regulated transcripts in the dark, we compared the activity of a *CAB* promoter fused to the *Escherichia coli* gene *uidA*, encoding β -glucuronidase (GUS), in light- and dark-grown dwarf and wild-type plants. The *CAB-uidA* fusion in pOCA107-2 (Li et al. (1994) *Genes Dev.*

25 8:339-349) was crossed into *dwf4*, and F2 dwarf and wild-type plants were grown in the dark or light for 12 days, followed by determination of GUS activity by fluorometry (Gallagher (1992). "Quantitation of GUS activity by fluorometry" in GUS Protocols, S.R. Gallagher, ed (New York: Academic Press), pp. 47-59).

The results demonstrated that when grown in the light, both wild-type and

30 *dwf4* seedlings contained GUS activity, which was significantly reduced in both lines when grown in the dark. Moreover, dark-grown *dwf4* seedlings displayed no GUS activity above the background present in dark-grown wild-type plants. The absence of

light-induced gene expression in the dark is a distinguishing feature of certain *cop* and *det* mutants, such as *cop2*, *cop3*, and *det3*. Because we have shown that the defect in cell elongation of *dwf4* is specifically rescued by BRs, even in the presence of light, we conclude that this is not a light-regulatory mutant. That its phenotype is partially
5 deetiolated or constitutively photomorphogenic is a secondary effect of its reduced stature and the growth conditions.

Example 8: Abnormal Skotomorphogenesis as a Consequence of the Dwarf Growth Habitat

10 When *dwf4* is grown in the light, its morphology is similar to that of various *cop* and *det* mutants, with multiple short stems, short rounded leaves, loss of fertility due to reduced stamen length, and delayed development (Figure 6). Dark-grown *dwf4* seedlings possess short hypocotyls, open cotyledons, and developing leaves. Therefore, it is tempting to speculate that this mutant may be defective in the control
15 of light-regulated processes. On the other hand, because a dark-flowering phenotype has been demonstrated for liquid-grown Arabidopsis (Araki and Komeda (1993), *supra*), and given that agar medium is mostly water, it is especially significant that it is the dwarf seedlings, whose apical meristems are very close to the agar surface, that display a light-grown phenotype in the dark. Furthermore, because wild-type
20 seedlings grown along the surface of the agar reproduce the dark-flowering phenotype, it is possible that the apparent light-regulatory defect of dwarf seedlings is a dark-flowering response. This possibility is strengthened by the observation that wild-type seedlings (ecotype Wassilewskija [Ws-2]) grown in the dark on horizontally oriented plates occasionally bend down and touch the agar surface, and these
25 seedlings invariably produce leaves.

In addition, of the eight *DWF* loci identified in this laboratory, only the shortest mutants displayed open cotyledons and leaf bud development; in the case of *dwf1* (Feldmann et al. (1989), *supra*), this aberrant skotomorphogenesis is confined to the most severely affected alleles. In addition to the presence of a short hypocotyl and
30 at least partially open cotyledons in the dark, *cop1* (Deng and Quail (1992), *supra*), *det1* (Chory et al. (1989b), *supra*), and *det3* (Cabrera y Poch et al. (1993) *Plant J.* 4:671-682) have been shown to initiate leaf formation in the dark. In mutants such as

cop1, *cop8*, *cop9*, *cop10*, and *cop11*, stomata undergo photomorphogenic maturation (Deng and Quail (1992), *supra*; Wei and Deng (1992), *supra*; Wei et al. (1994)), *supra*); of these, *cop1* and *cop9* as well as *det1* (Chory et al. (1989b), *supra*) also initiate differentiation of plastids into chloroplasts. *dwf4* displayed, in addition to a
5 light-grown dwarf phenotype, a dark-growth phenotype of short hypocotyls, open cotyledons, and developing leaves; however, in contrast with the light-regulatory defect seen with whole plants, the cellular differentiation phenotype was unaffected. In dark-grown dwarf seedlings, stomata did not complete their development, and differentiation of chloroplasts was not observed. The absence of a cellular
10 light-regulatory phenotype in *dwf4* is similar to that of a number of photomorphogenic mutants, such as *det2*, *det3*, *cop2*, *cop3*, and *cop4* (Chory et al. (1991), *supra*; Cabrera y Poch et al. (1993), *supra*; Hou et al. (1993), *supra*).

In view of the dark-flowering phenotype on agar and the absence of a light-regulatory defect in differentiating cells, we conclude that at least in the case of
15 *dwf4*, aberrant skotomorphogenesis may be a consequence of a dwarf growth habit rather than dwarfism being part of a defect in the control of light-regulated processes. This effect may also explain the light-regulatory phenotype found in other mutants with severely reduced height, such as *axr2* (Timpote et al. (1992), *supra*), and strong alleles of *dwf1*, both of which are also rescued by exogenous BRs (Szekeres et al.
20 (1996), *supra*).

Example 9: Feeding Experiments with BR Biosynthetic Intermediates

In view of the results described above, we hypothesized that DWF4 mediates one or more of several steroid hydroxylation steps in the BR biosynthetic pathway.
25 To test this, *dwf4* was grown on all of the available biosynthetic intermediates in the BR biosynthetic pathways and examined to ascertain which intermediates could rescue the dwarf phenotype. In addition to the intermediates belonging to the early C-6 oxidation and late C-6 oxidation pathways (Choi et al. (1997), *supra*), 22 α -hydroxycampesterol (22-OHCR), 6 α -hydroxycathasterone (6-OHCT) (Takatsuto et al. (1997) *J. Chem. Res. (synop.)* 11:418-419), and 6 α -hydroxycastasterone
30 (6-OHCS) (S. Takatsuto, T. Watanabe, T. Noguchi, and S. Fujioka, unpublished data) were synthesized and tested.

Germinated seedlings were transferred to media supplemented with one of the intermediates or BL to pinpoint the step catalyzed by DWF4. Cathasterone (CT; early C-6 oxidation pathway), 6-OHCT, 6-deoxocathasterone (6-deoxoCT; late C-6 oxidation pathway), and 22-OHCR, and all of the downstream compounds belonging to each branch, rescued the light-grown *dwf4* phenotype, whereas the known precursors failed to cause an elongation response. Rescued seedlings exhibited greatly elongated cotyledonary petioles and expanded cotyledons, moderately elongated hypocotyls, and leaves that were larger and not as curled compared with nonrescued dwarfs. In addition, the rescued seedlings were less green than the dwarfs. These experiments were conducted in liquid media. Feeding experiments performed in the dark yielded similar results.

Dose-response tests on the putative substrates and products of DWF4 were also performed. *dwf4* seedlings failed to respond to 6-oxocampestanol (6-oxoCN) even at high concentrations (3×10^{-6} M). However, on CT the overall morphology of *dwf4* was essentially rescued to wild-type phenotype at 3×10^{-7} M and higher, whereas with 6-deoxoCT, rescue occurred with as little as 10^{-7} M and may have even been inhibitory at higher concentrations. Of particular interest is the more dramatic response of the epicotyls versus the smaller response of the hypocotyls to CT. This same phenomenon was true for seedlings treated with $>10^{-7}$ M 6-deoxoCT. At concentrations $>10^{-7}$ M, the seedlings displayed an inhibition in hypocotyl and root elongation as well as cotyledon and leaf expansion.

In a dose-response experiment performed in the dark, the seedlings failed to respond to 6-oxoCN (10^{-8} to 3×10^{-6} M). A higher concentration of CT for dark-grown seedlings, compared with light-grown seedlings, 3×10^{-6} M (Figure 5B), was required to convert the hypocotyl to a length similar to that of the wild type. High concentrations of 6-deoxoCT caused dramatic elongation but were less effective at rescuing *dwf4* hypocotyls to wild-type phenotype.

To determine whether the results of the seedling feeding experiments could be applicable to soil-grown mature plants, 6-week-old *dwf4* plants were treated with BR intermediates and BL. Concentrations of applied intermediates were adjusted empirically to optimize responses. Consistent with the results obtained from the seedling experiments, only 22 α -hydroxylated compounds can rescue the *dwf4*

phenotype. The elongation response was only observed in the young tissues of the inflorescence, regardless of whether the BRs were applied locally or sprayed over the entire plant. In contrast to the striking elongation of the peduncles and pedicels, fertility was not restored by BR treatment. The sterility in *dwf4* is hypothesized to be
5 mechanical, which means that the filaments are shorter than the carpels such that the pollen is shed onto the ovary walls rather than onto the stigmatic surface. In fact, if *dwf4* plants are hand pollinated using *dwf4* pollen, fertility increases.

Pedicels displayed a more consistent response to exogenously applied BRs than did internodes, which led us to quantify the sensitivity of pedicels to these
10 compounds. As shown in Figure 7, *dwf4* pedicels were more sensitive to BR intermediates belonging to the late C-6 oxidation pathway, 6-deoxoCT (10^{-6} M) and 6-deoxoteasterone (6-deoxoTE; 10^{-6} M), compared with CT (10^{-5} M) and teasterone (TE; 10^{-5} M) of the early C-6 oxidation pathway. The end product of the BR pathway, BL (10^{-7} M), possessed the highest bioactivity. This concentration induced
15 approximately the same degree of response as its precursor compounds at 10^{-6} M. Finally, application of 22-OHCR (10^{-5} M) also resulted in a dramatic elongation response (Figure 7).

Rescue of *dwf4* by 22-hydroxylated steroids confirms that the missing step in *dwf4* is hydroxylation at the C-22 position. In fact, we found that the chemically
20 synthesized 22-OHCR was also effective in rescuing *dwf4* (Figure 7) hydroxylation at C-22. These results indicate that there is no defect other than 22 α -hydroxylation in *dwf4* plants.

In BR biosynthesis, Fujioka and Sakurai (1997b), *supra* have demonstrated that there are at least two branched biochemical pathways to the end product BL
25 (Figure 1; Fujioka and Sakurai (1997a), *supra*, Fujioka and Sakurai (1997b), *supra*; Sakurai and Fujioka (1997), *supra*). Depending on the oxidation state of C-6, they are referred to as the early or late C-6 oxidation pathways. In the early pathway, the C-6 is oxidized to a ketone at campestanol (CN), whereas in the late pathway it is oxidized at 6-deoxocasterone (6-deoxoCS). Otherwise, the two pathways share equivalent
30 reactions. Our results from the experiments with the available BR intermediates clearly demonstrate that *dwf4* is defective in the 22 α -hydroxylation steps in each of the pathways. Application of all 22 α -hydroxylated intermediates in these pathways,

such as CT and 6-deoxoCT, cause dramatic elongation of *dwf4* plants, but compounds *not* hydroxylated at C-22 had no effect. This result also suggests that DWF4 recognizes at least two substrates: CN and 6-oxoCN. It seems reasonable to hypothesize that the same result will be found for *CPD*, a 23 α -hydroxylase; that is, it
5 will use 6-deoxoCT as well as CT as substrate.

The rescue of *dwf4* by 22-OHCR is an important observation. First, it confirms DWF4 as a 22 α -hydroxylase. Second, this result suggests that 22-OHCR was metabolized to induce the same responses as other complementing BRs. This is not just a general effect because our unpublished data show that another dwarf mutant
10 that we have identified in our screens, *dwf8-1*, is not rescued by this compound. Finally, these feeding experiments suggest that the metabolism of 22-OHCR may represent a new subpathway in the BR biosynthetic pathway. If this compound also exists *in vivo* and constitutes the first step in a separate subpathway, by analogy to the chemical structure, the C-6 hydroxylated BRs, for example, 6-OHCT,
15 6-hydroxyteasterone, and so on, may be possible intermediates in this network. If so, the intermediates in this pathway may play a role as bridging molecules between the early and late C-6 oxidation pathways. Alternatively, it might be possible that 22-OHCR merges into one of the two pathways to be metabolized. In this case, the late C-6 oxidation pathway is the best candidate; our unpublished data show that
20 22-OHCR is more effective in the light in rescuing the *dwf4* phenotype, which is true for all of the intermediates in the late C-6 oxidation pathway.

Currently, biochemical feeding studies suggest that the two pathways merge to produce BL or CS (Yokota et al. (1991), Metabolism and biosynthesis of brassinosteroids. In *Brassinosteroids: Chemistry, Bioactivity, and Application*, H.G. Cutler, T. Yokota, and G. Adam, eds (Washington, DC: American Chemical Society),
25 pp. 86-96; Yokota, et al. (1997) *Plant Physiol.* 115(suppl.):169; Figure 1). Several lines of evidence indicate that seemingly redundant pathways can be utilized to respond to environmental or developmental signals. First, the pathways could respond to specific signals. For instance, it is possible that various cues such as light, dark, or developmental signals play a role in regulating these subpathways. Our
30 feeding experiments consistently showed that BRs in the late C-6 oxidation pathway are more effective at promoting cell elongation in light-grown plants (*dwf4* and wild

type; Figure 7) and that the BRs belonging to the early C-6 oxidation pathway are more active in dark-grown seedlings. Thus, it may be possible that the late C-6 oxidation pathway operates in the light and that the early C-6 oxidation pathway functions primarily in the dark. Second, rather than a simple merger of branched pathways to BL as an end product, each intermediate may have nascent bioactivity. The in vivo ratio or composition of BRs at different oxidation states may result in different responses. Noticeably distinctive phenotypes for the various BR dwarfs, defective in different biosynthetic steps, support this idea. Third, the biosynthetic rate of each pathway toward production of the end product may differ. In this case, the biosynthetic rate could be modulated by controlling the level of gene expression or the activity of participating enzymes. Certain signals, requiring different rates of BR biosynthesis, may induce one of the subpathways, which would then affect the concentration of the intermediates in one pathway relative to the other.

Of the steps in BR biosynthesis in Madagascar periwinkle, the 22α -hydroxylation reaction has been suggested to be the rate-limiting step (Fujioka et al. (1995a) *Biosci. Biotech. Biochem.* 59:1543-1547). In periwinkle, the endogenous level of CT was as low as one-twenty thousandth of CR; however, CT was almost 500 times more active than 6-oxoCN in the rice-lamina inclination assay (Fujioka et al. (1995b) *Biosci. Biotech. Biochem.* 59:1973-1975). Based on these results, we propose that the step encoded by *DWF4* serves as the rate-limiting reaction and that once past this step, the intermediates are easily converted to the end product. Although biochemical studies on *DWF4* need to be performed to ascertain whether it mediates the rate-limiting step, *DWF4* seems to be greatly downregulated compared with *CPD*, the next enzyme in the pathway; RT-PCR revealed that the *DWF4* transcript is much less abundant than the *CPD* transcript.

Example 11:

A. Promoter and overexpression constructs

Two promoter constructs were used for the *DWF4-promoter::GUS* (*D4G*) analysis. For promoter fusions, polymerase chain reaction (PCR) products spanning 1.1 kb DNA upstream of the translation initiation site were amplified using primers

D4XLINIT (5'-TAGGATCCAGCTAGTTTCTCTCTCTCT-3') (SEQ ID NO:16) and a T7 primer (5'-TAATACGACTCACTATAGGG-3') (SEQ ID NO:17). For template for PCR, a *DWF4* genomic clone subcloned into pBluescript SK- vector (Stratagene, La Jolla, CA) was used as described herein. The PCR products were
5 restricted with *SaII* and *BamHI*, and ligated into the same restriction site of a promoterless GUS vector pBI101; this 1.1 kb *promoter::GUS* construct was named pD4GL. For the pD4GS construct, pD4GL was digested with *HindIII*, the small restriction fragment was removed, and the remaining vector with the partial promoter was self-ligated. The constructs were introduced into *Agrobacterium* strain GV3101
10 through electroporation.

For a *DWF4* overexpression construct, PCR products were made by using D4OVERFA (5'-GAATTCTAGAATGTTTCGAAACAGAGCATCATA-3') (SEQ ID NO:18) and D4R2 (5'-CCGAACATCTTTGAGTGCTT-3') (SEQ ID NO:10) primers and Wassilewskija-2 (Ws-2) genomic DNA. The PCR products were cut with *XbaI*
15 and *HindIII*, and inserted into the same restriction sites of genomic clone SCH25 containing a 2.5 kb *HindIII* fragment of the *DWF4* DNA corresponding to the 3' half of the gene. The resulting recombinant DNA clone pD4CDS, containing the whole coding sequence from the translation initiation site to 694 bp downstream of the stop codon, was cut with *XbaI* and transferred to an overexpression vector pART27
20 (Gleave (1992), *Plant Molec. Bio.* 20:1203-1207). The resulting binary construct was named pOD4. This construct was introduced into *Agrobacterium* through electroporation.

B. Spray transformation

25 Since it has been shown that *Agrobacterium*-mediated transformation can work by seed infection (Feldmann and Marks (1987) *Molec. Gen. Genet.* 208:1-9) or by simply dipping the host plants into *Agrobacterium* culture, we decided to try spraying the *Agrobacterium* directly onto the plants. In addition to spraying, the "floral dip" method was used as described (Clough and Bent (1998), *infra*). About
30 Wassilewskija-2 (Ws-2) wild-type and *dwf4-4* seeds were sprinkled on 10 cm pots, and thinned to 5-6 plants per pot 10 days (wild type) and 20 days (*dwf4-4*) after germination. When the primary inflorescences of the wild type reached 3-4 cm in

height, they were decapitated to induce axillary bolts. *dwf4-4* plants were used without decapitation. For the preparation of *Agrobacterium*, a single colony selected on 20 $\mu\text{g/ml}$ kanamycin in Luria-Bertani (LB) medium (10 g bacto-tryptone, 5 g bacto-yeast extract, 10 g NaCl per liter, pH 7) was inoculated into 100 ml liquid LB media, and
5 grown for 3 days. One OD_{600} unit equivalent cells were used to inoculate 100 ml LB media. The overnight grown cells were collected by centrifugation, and resuspended with transformation media as described in Clough and Bent (1998), *infra* (5% sucrose and 0.05% Silwet L-77, $\text{OD}_{600} = 1$). The *Agrobacterium* suspension was sprayed onto plants on the third day after decapitation. To avoid physical contact with possibly
10 hazardous Silwet vapor, protective glasses were used and the spraying was done in a fume hood. To test the efficiency of repeated spraying, plants were sprayed every third day (3x). Sprayed plants were grown to maturity and seeds harvested. For seed sterilization 0.07 g seeds were surface sterilized by treating for 2 min in 70% ethanol, 15 min in bleach solution consisting of 5% Clorox and 1% SDS, followed by three
15 rinses with sterile water. To plate the seeds 25 ml of sterile top agar (0.15% agar) was added to the sterilized seeds and the seed mixture was poured onto Murashige and Skoog solid plate (100 x 15 mm, Murashige and Skoog salts, 5% sucrose, 0.08% agar, pH 6) supplemented with kanamycin or hygromycin at 60 $\mu\text{g/ml}$ and 40 $\mu\text{g/ml}$, respectively. Twelve days after germination kanamycin resistant were transferred to
20 single pots, and grown to maturity. T2 seeds were collected from individual transformants (T1), and plated again on the selection media to determine segregation ratios for drug-resistant versus sensitive plants. *Arabidopsis* transformants were named *Arabidopsis* Qverexpressor of DWF4 (*AOD4*) when harboring an overexpression construct pOD4, and DWF4-*promoter*::GUS (*D4G*) for transformants
25 containing a GUS fusion gene. Homozygosity for the transgene was determined when no sensitive T4 seedlings segregated from >500 T3 individuals. Morphometric analysis of *AOD4* lines and GUS histochemical analysis of *D4GL* plants was performed using plants homozygous for the transgene.

For histochemical analysis of the *D4GL* plants, seeds were plated on M&S
30 plates and grown in the dark and light. Seedlings were harvested at the designated dates and stained overnight using a substrate mixture (0.1 M NaPO_4 , pH 7, 10 mM EDTA, 0.5 mM $\text{K}_3\text{Fe}(\text{CN})_6$, 0.5 mM $\text{K}_4\text{Fe}(\text{CN})_6$, 1 mM X-glucuronide, and 0.1%

Triton X-100). Seedlings cleared with 90% ethanol were rehydrated before taking pictures using a Stemi SV11 dissecting microscope (Zeiss, NY).

Transgenic tobacco plants (*TOD4*) harboring the pOD4 constructs were produced in the plant tissue culture laboratory at the University of Arizona. Protocols
5 for the regeneration of transgenic plants from leaf discs of *Nicotiana tabacum* var Samsun will be provided on request. Fifteen independent transformants for both the control and OD4 constructs were grown for seeds. Morphological analysis of the *TOD4* lines was performed using T2 plants in the course of growth for 4 months in the green house (30°C). Methods for *Arabidopsis* growth and RNA gel blot analysis
10 were previously described herein. Briefly, seeds of wild type and the two *AOD4* lines were germinated on M&S agar media. 10 days after germination, 20 seedlings confirmed to be resistant to kanamycin were transferred to a single pot. Various morphological traits (Table 3) were measured. To determine the seed production, after 8 weeks from germination, plants were further dried for two weeks at room
15 temperature. Seeds were harvested from an individual plant and weighed. To measure the seed size, seeds were magnified 3 times under the dissecting microscope, the width and the length of five seeds from each plant were measured to the nearest tenth of mm.

20 **C. *DWF4* transcription is localized to zones of cell division and elongation**

To localize BR biosynthesis, RNA gel blot analysis with total RNA isolated from nine different tissues of three-week old plants was performed. The *DWF4* transcript was barely detectable in shoot tips, roots, dark-grown seedlings, callus and axillary buds, but the levels were below the detectable limit in the other tissues
25 examined, including stems, siliques, pedicels, and rosette leaves. For finer localization of the expression, the expression of the GUS reporter gene controlled by the *DWF4* promoter was examined.

Prior to performing *DWF4-promoter::GUS* gene fusion analysis, a 1.1 kb fragment of DNA upstream of the *DWF4* translation start site was tested to ensure that
30 it contained all of the necessary sequence elements for proper transcriptional control of *DWF4*. *dwf4-4* plants were transformed with a 4.8 kb construct consisting of a 1.1 kb promoter region and 3.7 kb that contained the complete *DWF4* coding sequence.

For *Agrobacterium*-mediated transformation of *Arabidopsis* plants, a "spray transformation" protocol was employed rather than traditional methods. Spray transformation yielded a comparable number of transformants relative to the traditional "floral dip" (Clough and Bent (1998) *Plant J.* 16:735-743) or "vacuum infiltration" methods (Bechtold et al. (1998) *Methods Mol. Biol.* 82:259-266).
5 Interestingly, repeated spraying resulted in an increased number of transformants. Transformants harboring the *DWF4* genomic DNA displayed a wild type phenotype, suggesting that the promoter segment contained the necessary information for proper expression of the gene.

10 For histochemical staining analyses of transgenic plants harboring the *DWF4*-*promoter::GUS* (*D4G*) recombinant gene, two different *D4G* constructs were made and tested. *D4GL* contained the 1.1 kb promoter fragment, whereas *D4GS* carried only a TATA-like promoter region (280 bp). GUS staining in 20 independent transformants containing *D4GS* was either not detected or inconsistent between
15 transformants. However, the 20 transgenic plants containing *D4GL* displayed a consistent GUS staining pattern, suggesting that the 1.1 kb promoter is required for the proper transcriptional control of *DWF4*.

Analyses of GUS staining patterns in T2 plants homozygous for *D4GL* revealed that GUS activity was present in tissues with actively dividing or elongating
20 cells. These include shoot apical meristems, leaf primordia, collet (the junction between hypocotyl and root), and root tips, including lateral root primordia, as shown in 6-day old light-grown seedlings. Interestingly, dark-grown seedlings displayed GUS activity in cotyledons whereas the staining was not detectable in the cotyledons of light-grown seedlings. In adult plants, GUS activity was detected in floral
25 primordia, carpels, and the basal end of the filaments of unopened flowers, whereas GUS activity in sepals, petals, and mature pedicels was not detected. The shoot tips, bases of emerging branches, and primordia of axillary inflorescences were GUS positive, whereas elongated internodes were negative. Embryos in the seeds of the fully elongated siliques were weakly positive for GUS staining, suggesting a role for
30 BRs in embryo development. Leaf primordia, young leaves, expanding leaf margins, and the base of petioles displayed GUS activity, but old leaf blades were negative for GUS staining. The tissues positive for GUS staining confirmed the expression pattern

examined by northern analysis with the tissue-specific RNA.

Since DWF4 is proposed to be a key enzyme in the BR biosynthetic pathway, *DWF4* transcription could be regulated by an end-product feedback mechanism. To this end, *D4GL* was expressed in different genetic backgrounds including two BR
5 deficient mutants, *dwf7-1* and *dwf8-1*, and a BR-enriched line, *AOD4*. GUS activity was increased in *dwf7-1* and *dwf8-1* but decreased in *AOD4* lines. DWF7 is a C-5 desaturase that acts in the sterol specific part of the pathway. *D4GL* activity in *dwf7-1* was found in the same tissues as wild type but. *dwf8-1* is defective in a BR biosynthetic step downstream of *CPD*. In *dwf8-1*, the intensity of the *D4GL* activity
10 was noticeably stronger as compared to wild type but the expression patterns were relatively diffuse. *dwf8-1* was also found to express GUS at nascent sites as compared to wild type. In wild type, *D4GL* expression in the cotyledons of light-grown seedlings was not detected, but *dwf8-1* displayed considerable *D4GL* activity in the cotyledons. Also in contrast to wild type, GUS activity was detected throughout the
15 hypocotyls of *dwf8-1* light-grown seedlings, suggesting that *D4GL* transcription is upregulated in *dwf8-1* in a more general manner. Conversely, GUS activity was greatly reduced in *AOD4-4* plants. Also, in *AOD4* plants GUS activity in the root tip and collet was completely eliminated, whereas the shoot tip retained residual activity, suggesting that increased levels of BRs in *AOD4-4* may have resulted in lower GUS
20 activity. The down-regulation of GUS activity was similarly found if *D4GL* plants were exogenously supplied with 10^{-6} M 24-epibrassinolide (epi-BL). Seedlings treated with epi-BL displayed greatly reduced GUS activity in tissues normally stained in untreated control plants, suggesting that exogenously applied epi-BL effectively down-regulates *D4GL* activity. However, hypocotyls of *D4GL* plants
25 supplemented with 10^{-4} M GA_3 , while longer than controls, did not display an increase in GUS staining in shoots and roots. This suggests that GA_3 or GA_3 -induced elongation did not affect *D4GL* transcription in these tissues.

D. DWF4 overexpression results in elongated hypocotyls in *Arabidopsis* and tobacco seedlings

A *DWF4* overexpression construct (pOD4) was made by placing the *DWF4* genomic DNA under the control of the CaMV 35S promoter. RNA gel blot analysis,

with total RNA isolated from the transgenic lines containing the overexpression construct, showed that *DWF4* transcripts were greatly increased in both *Arabidopsis* and tobacco, whereas the level was not readily detectable in either wild type or in *dwf4-1* plants. Similar to increased mRNA transcripts, the 80 independent *AOD4* transgenic plants had longer hypocotyls and inflorescences.

To compare the phenotypic effects resulting from the endogenous and exogenous addition of BRs, the length of roots and hypocotyls of 16 seedlings of *dwf4*, wild-type controls, wild-type plants supplemented with 10^{-6} M epi-BL, and two independent *AOD4* lines, grown for 12 days in the light or dark was measured. As described herein, *dwf4-1* displayed greatly reduced hypocotyl length both in the light and dark as compared to wild type. Wild-type roots are shortened when grown in the dark, but *dwf4-1* root length was not significantly reduced in the dark compared with the reduction in hypocotyl length. When epi-BL is added, light-grown wild type seedlings developed elongated hypocotyls, whereas roots were shorter than untreated control plants. These characteristic responses of wild-type plants to epi-BL treatment were similar in two independent *AOD4* lines. The hypocotyl length of light-grown *AOD4* seedlings was comparable to that of seedlings treated exogenously with epi-BL. However, dark-grown hypocotyls showed a dramatic increase in length as compared to controls with or without epi-BL. Inhibition of root growth was also obvious in the *AOD4* lines. Furthermore, the increased hypocotyl length and reduced root length were consistently observed in 15 independent transformants of tobacco (*TOD4*) harboring a pOD4 construct. This result suggests that the *Arabidopsis* DWF4 enzyme also catalyzes BR biosynthesis in tobacco.

E. DWF4 overexpression results in increased plant height, bigger leaves, and increased seed production

As shown below in Table 3, the effects of *DWF4* overexpression on plant growth were monitored during the course of development. The number of rosette leaves at bolting was not significantly different between wild-type and *AOD4* plants (Table 3). The inflorescence height of wild type and two independent *AOD4* lines were comparable 20 days after germination (DAG). Later, the *AOD4* lines outgrow wild type. Surprisingly, *AOD4* lines continue to grow beyond 35 DAG at the time

wild-type plants ceased elongation. At maturity, the height of *AOD4* lines was 135% (*AOD4-65*) and 142% (*AOD4-73*) that of wild type, respectively. Similarly, *TOD4* also displayed a 14% increase in plant height as compared to the control.

5 Interestingly, the increased inflorescence length in *AOD4* plants seemed to be at the cost of stem stiffness. During development, *AOD4* plants tend to fall over earlier than the *Ws-2* wild type. In addition to plant height, comparison of rosette leaf size between wild type and *AOD4* indicates that leaves, both rosette and cauline, are larger, especially in adult plants. *TOD4* plants also possessed leaves that were larger, and had longer petioles relative to the control. Furthermore, additional secondary
10 branches were found both in *Arabidopsis* and tobacco overexpression lines. In *AOD4* plants, this additional branching was associated with >2 times increased number of siliques per plant, leading to a 33 and 59% increase in seed production (Table 3). The increased seed production in the *AOD4* lines was mainly due to the increased number of seeds per plant than increase in the seed size, because the size was not significantly
15 increased (Table 3). In addition to the increased number of seeds, the length of silique as well as the length of an internode between the first silique in a main inflorescence and the base of plant was increased (Table 3).

Figure 8 shows that stem growth is increased more than 20% compared to wild type in DWF4 overexpression lines and Figure 9 shows that seed production is
20 increased significantly over wild type in the DWF4-overexpressed lines. Figure 12 depicts hypocotyl length and root length in light and dark. Further, the height of *AOD4* lines was greater than wild type over the days examined. In addition, although wild type plants ceased growth around five weeks after germination, *AOD4* plants continued to grow up to seven weeks.

5
10

Table 3. Morphological comparison among control and AOD4 lines.

	Ws-2	AOD4-65	AOD4-73	Percent increase over Ws-2	
				AOD4-65	AOD4-73
Size					
Height in cm ¹	11.4 ± 1.8	15.4 ± 3.3	16.8 ± 3.8	135	147
Silique in mm	13.7 ± 2.0	18.4 ± 2.5	20.0 ± 3.2	134	145
Seed length in mm	0.47 ± 0.04	0.51 ± 0.05	0.49 ± 0.03	108	105
Seed width in mm	0.29 ± 0.03	0.32 ± 0.03	0.3 ± 0.03	110	102
Number					
Rosette	5.1 ± 0.5	5.2 ± 0.4	5.7 ± 0.5	100	112
Branch ²	9 ± 3	23.4 ± 5.7	21.6 ± 6.8	260	240
Siliques	106.7 ± 33.6	373.5 ± 107.5	344.2 ± 47.1	350	325
Seed production ³					
(mg/plant)	164 ± 48	218 ± 33	261 ± 37	133	159

¹ Between rosette and the first silique (N>12)² Number of branches bearing siliques (N>12)³ Weight of seeds per individual plant (N>12)

Thus, the *DWF4* locus is defined by at least four mutant alleles. One of these is the result of a T-DNA insertion. Plant DNA flanking the insertion site was cloned and used as a probe to isolate the entire *DWF4* gene. Sequence analysis revealed that *DWF4* encodes a cytochrome P450 monooxygenase with 43% identity to the putative

5 Arabidopsis steroid hydroxylating enzyme CONSTITUTIVE
PHOTOMORPHOGENESIS AND DWARFISM. Sequence analysis of two other mutant alleles revealed deletions or a premature stop codon, confirming that *DWF4* had been cloned. This sequence similarity suggests that *DWF4* functions in specific hydroxylation steps during BR biosynthesis. The dwarf phenotype can be rescued

10 with exogenously supplied brassinolide. *dwf4* mutants display features of light-regulatory mutants, but the dwarfed phenotype is entirely and specifically brassinosteroid dependent; no other hormone can rescue *dwf4* to a wild-type phenotype. Feeding studies utilizing BR intermediates showed that only

15 22 α -hydroxylated BRs rescued the *dwf4* phenotype, confirming that *DWF4* acts as a 22 α -hydroxylase. In adult plants, strong GUS staining (indicative of *dwf4* expression) was found in the primordia of axillary inflorescences and secondary branches, and in young developing flowers. GUS expressing tissues correspond to the tissues sensitive to exogenously applied BRs leading to the hypothesis that these tissues are putative brassinolide biosynthetic sites. The inflorescence height of *DWF4* overexpressing

20 lines increased >35% in *Arabidopsis* (*AOD4*) and 14% in tobacco (*TOD4*) as compared to control plants at maturity. The total number of branches and siliques increased >2-fold in *AOD4* plants, leading up to a 59% increase in seed production. The phenotypes of *dwf4*, *DWF4*, and *AOD4* plants suggest that the degree of *DWF4* transcription is associated with the degree of BR effects. In sum, it appears be

25 possible to engineer agricultural plants with increased biomass and seed yield.